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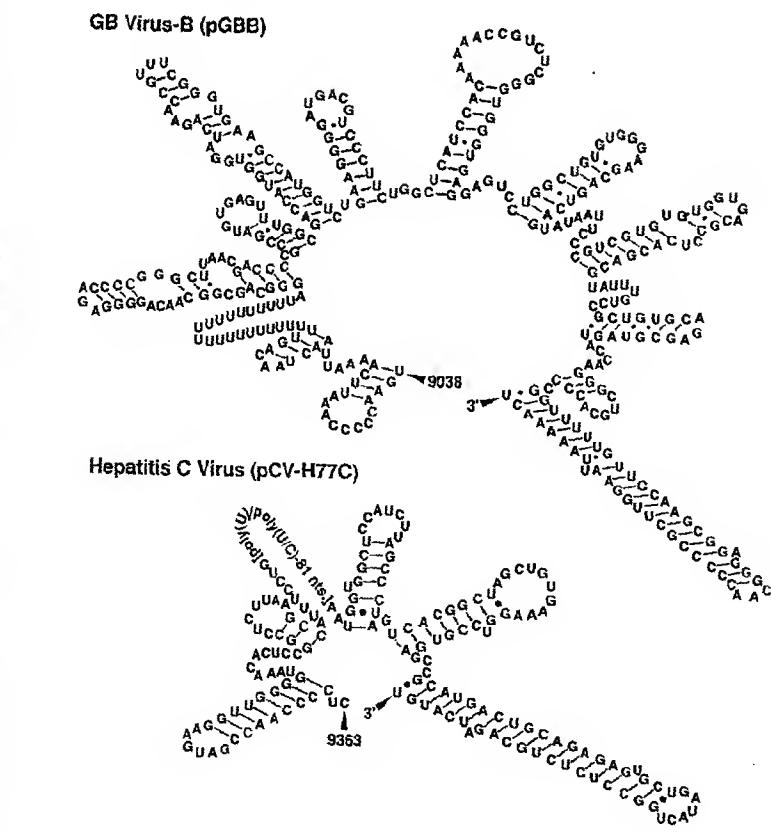
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(54) Title: INFECTIOUS cDNA CLONE OF GB VIRUS B AND USES THEREOF



(57) Abstract: The present invention relates to nucleic acid sequence which comprises the genome of an infectious GB virus B clone. The invention also relates to the use of the nucleic acid sequence of the infectious GB virus B clone to indirectly study the molecular properties of HCV, and in the production of HCV/GBV-B chimeras. The invention further relates to the use of the infectious nucleic acid sequence of GB virus B clone and the HCV/GBV-B chimeras in the development of vaccines and therapeutics for HCV.



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◦ Title of Invention

Infectious cDNA clone of GB Virus B and Uses Thereof

Field of Invention

5 The present invention relates to nucleic acid sequence which comprises the genome of an infectious GB virus B (GBV-B) clone. The invention also relates to the use of the nucleic acid sequence of the infectious GB virus B clone to study indirectly the molecular
10 properties of hepatitis C virus (HCV), and in the production of HCV/GBV-B chimeras. The invention further relates to the use of the infectious nucleic acid sequence of the GB virus B clone and the HCV/GBV-B chimeras in the development of vaccines and therapeutics
15 for HCV.

Background of Invention

20 Transmission studies of potential human hepatitis agents were first reported in 1967 (Deinhardt 1967). Four tamarins inoculated with acute phase sera from a surgeon with acute hepatitis (patient GB) developed hepatitis, as did most tamarins inoculated in serial passage studies. Subsequent studies indicated
25 that the etiological agent responsible for the development of hepatitis in these animals was not any of the known human hepatitis viruses (Purcell 1994). In 1995, two related RNA viruses named GB virus-B (GBV-B) and GB virus A (GBV-A) were identified in acute phase sera of a tamarin which developed hepatitis following inoculation with serum of the eleventh tamarin passage of the putative GB agent (Simons 1995a).

30

35 GBV-B infection of tamarins resulted in acute resolving hepatitis (Schlauder 1995, Buhk 1997). The

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- natural host of GBV-B is still unknown as the virus has not been detected in uninoculated animals or in humans.

5 GBV-A, on the other hand, is an indigenous tamarin virus rather than a component of the original GB inoculum (Bukh 1997, Erker 1998). Experimental infection of tamarins with GBV-A did not produce hepatitis (Schlauder 1995). A human agent, GBV-C or hepatitis G virus, most closely related to GBV-A, was later identified (Simons 1995b, Linnen 1996). However, 10 it is still not clear whether this virus actually causes hepatitis (Alter 1998, Bukh 1998a). Thus, of the known GB viruses, GBV-B may be the only true hepatitis virus.

15 Based on analysis of their genomic sequences, GBV-A, GBV-B and GBV-C were classified as members of the *Flaviviridae* family of viruses, and among the known viruses, GBV-B is the virus most closely related to hepatitis C virus (HCV) (Muerhoff 1995, Robertson 1998).

20 The GBV-B virus contains a positive-sense, single-stranded RNA genome of 9143 nucleotides (nts) (Simons 1995a, Muerhoff 1995). The viral genome of GBV-B consists of a 5' untranslated region (UTR), a single long open reading frame (ORF) and a 3' UTR. Based on 25 known motifs, structural proteins were predicted to be encoded in the 5' portion of the ORF and nonstructural (NS) proteins in the 3' portion of the ORF (Muerhoff 1995). The hydropathy plots of the polyproteins of GBV-B and HCV are very similar even though the overall 30 homology of the predicted polyproteins between GBV-B and HCV is only about 25-30% (Muerhoff 1995). The putative envelope proteins (E1 and E2) of GBV-B and HCV share common structural features, and significant homology was 35 observed between the NS3 serine protease, the NS3 RNA

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o helicase, and the NS5 RNA-dependent RNA polymerase regions of GBV-B and HCV (Muerhoff 1995). Furthermore, the function and substrate specificity of the GBV-B and HCV NS3 serine proteases are also similar (Scarselli 1997). The genomic structure and organization of GBV-B and HCV share additional features of interest. First, colinear regions with significant sequence homology were identified in the 5' UTRs (Muerhoff 1995) and the predicted IRES structure of GBV-B is similar to that of HCV (Lemon 1997). Second, both viruses begin the 3' UTR with a short sequence followed by a poly (U) stretch followed by additional nucleotides (50 nucleotides for GBV-B and 98 nucleotides for HCV). However, the 3' terminal sequence of HCV forms a stable stem-loop structure (Kolykhalov 1996) whereas the published 3' terminal sequence of GBV-B does not.

To date, molecular studies of HCV are severely limited by the lack of an efficient cell culture system for the virus and by expense and limited availability of chimpanzees, the sole animal model for HCV. Accordingly, a less expensive and more readily available animal than chimpanzees is necessary as an animal model for the study of HCV.

Summary of Invention

The present invention relates to nucleic acid sequence which comprises the genome of an infectious GB virus B (GBV-B) clone. It is therefore an object of the invention to provide nucleic acid sequence which encodes an infectious GBV-B. Such nucleic acid sequence is referred to throughout the application as "infectious nucleic acid sequence".

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o As significant structural homology exists between the genomes of GBV-B and HCV, the invention also relates to the use of infection of tamarins with the infectious nucleic acid sequence of GBV-B or with 5 mutants of the infectious sequence to study indirectly the molecular properties of hepatitis C virus (HCV) or as a preliminary screen to identify agents which have antiviral activity against HCV.

10 The invention further relates to "chimeric nucleic acid sequences" consisting of portions of the infectious nucleic acid sequence of GBV-B and portions of the nucleic acid sequences of other viruses closely related to GBV-B such as HCV, GBV-C or other members of 15 the *Flaviviridae* family which do not replicate in tamarins. In a preferred embodiment, the chimeric nucleic acid sequences consist of portions of the infectious nucleic acid sequence of GBV-B and portions of the nucleic acid sequence of HCV. The nucleic acid 20 sequences taken from GBV-B and HCV can be open-reading frame sequences, and/or sequences from the 5'UTR and/or 3'UTR.

25 In one embodiment, GBV-B/HCV chimeras may be made in which 5' or 3' UTR sequences of the GBV-B infectious clone are replaced with the corresponding sequence from an HCV clone.

30 In another embodiment, GBV-B/HCV chimeras may be constructed in which the structural or non-structural regions of GBV-B are replaced by corresponding regions of HCV. Thus, such a chimera would contain, for example, the HCV structural region in a GBV-B "genomic backbone". Of course, it is understood by one of skill 35 in the art that the construction of the above-described

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- o chimeric nucleic acid sequences may be reversed such that, for example, the GBV structural region may replace the structural region of an HCV genome to produce a chimera in which the GBV structural region is contained in an HCV backbone.

5 The invention further relates to the use of the chimeric nucleic acid sequences of the invention to study the functions of HCV genes, and for the development of vaccine and antiviral agents against HCV.

10 The invention also relates to the use of the infectious GBV-B nucleic acid sequence, the mutated GBV-B nucleic acid sequences or the chimeric sequences of the invention to identify cell lines capable of supporting the replication of GBV-B or the chimeras of the invention.

15 The present invention also relates to the polypeptides encoded by the nucleic acid sequences of the invention or fragments thereof.

20 The present invention further relates to the in vitro and in vivo production of GBV-B, mutant GBV-B viruses or chimeric GBV-B/HCV viruses from the nucleic acid sequences of the invention.

25 The invention also provides that the nucleic acid sequences and viruses of the invention be supplied in the form of a kit, alone or in the form of a pharmaceutical composition.

30 Brief Description Of Figures

Figure 1 shows a flow diagram of GB virus transmission studies in two species of tamarins, *Saguinus mystax* (SM) and *Saguinus oedipus* (SO). The animals infected with GBV-B (Simons 1995a) are boxed.

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- Two serum pools (GB 8/93 and GB 2/94) were made from acutely infected animals. Both pools contained GBV-B, as well as GBV-A (Simons 1995) at a titer of 10^8 genome equivalent (GE)/ml. A 10% liver homogenate (CT 11/91) was made from a sacrificed tamarin. A number of *S. mystax* tamarins (SM 737, 749, 750, 760, 782, 795 and 799) and *S. oedipus* tamarins (SO 100) were naturally infected with GBV-A_{SM} and GBV-A_{SO}, respectively, prior to inoculation (Bukh 1997). Only two tamarins (SM 720 and 748), both GBV-A_{SM} negative, became infected with GBV-A (Simons 1995) following inoculation. Tamarins SM42 and SM670 were not tested for GBV-A or GBV-A_{SM}.

Figure 2 shows the course of GBV-B infection in tamarins (*S. mystax*) inoculated with a dilution series of the GB 2/94 pool. All animals were inoculated intravenously at week 0 with 1 ml of the indicated dilution. Results of qualitative RT-nested PCR for GBV-B in serum are shown at the top (filled circles, positive; empty circles, negative). Serum levels of isocitrate dehydrogenase (ICD in units/ml); shaded area) and the estimated \log_{10} GBV-B GE titer (vertical columns) were plotted against time.

Figure 3 shows alignment of the 3' UTR sequences of GBV-B. The sequence of the infectious clone of GBV-B (pGBB) is shown at the top (nts. 9038-9399). The other sequences shown are: pGBB5-1, a non-infectious clone of GBV-B; GBV-B, a prototype of GBV-B (Simons 1995); eleven "gb" clones obtained from CT 11/91 liver homogenate by 5' RACE on the minus-strand GBV-B RNA; four "29" clones obtained from GB 2/94 pool by RT-PCR across 5'-to-3'-end-ligated viral GBV-B RNA; and seven "GBB3" clones obtained from GB 2/94 pool by standard RT-PCR.

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- ° With pGBB as the reference, nucleotide substitutions or insertions are shown as uppercase letters, identical nucleotides are shown as dots and nucleotide deletions are shown as dashes.

5 Figure 4 shows the predicted secondary structure of the 3' UTRs of GBV-B and HCV as determined by the program "mfold" (Genetics Computer Group).

10 Figure 5 shows the course of GBV-B infection in *S. mystax* tamarins transfected with RNA transcripts of pGBB. Both animals were negative for GBV-A_{SM}. At week 0 transcription mixtures were injected into tamarins by percutaneous intrahepatic injection guided by ultrasound. Results of qualitative RT-nested PCR for GBV-B in serum 15 is shown at the top (filled circles, positive; empty circles, negative). Serum levels of isocitrate dehydrogenase (ICD in units/ml; shaded area) and the estimated log₁₀ GBV-B GE titer (vertical columns) were plotted against time.

20 Figures 6A-6F show the nucleotide sequence of the infectious hepatitis C virus clone of genotype 1a strain H77C and Figures 6G-6H show the amino acid sequence encoded by the clone.

25 Figures 7A-7F show the nucleotide sequence of the infectious hepatitis C virus clone of genotype 1b strain HC-J4 and Figures 7G-H show the amino acid sequence encoded by the clone.

30 Description of The Invention

The present invention relates to nucleic acid sequence which comprises the genome of an infectious GB virus B (GBV-B) clone. The nucleic acid sequence which 35 comprises the genome of an infectious GBV-B virus is

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- ° shown in SEQ ID NO:1 and is contained in the plasmid construct pGBB deposited with the American Type Culture Collection (ATCC) on May 28, 1999 and having ATCC accession number PTA-152. The present invention relates 5 to the identification of a 260 nucleotide sequence at the 3' end of the infectious GBV-B clone which is shown in Example 3 to be necessary for the development of the infectious clone.

Since GBV-B is the virus most closely related 10 to HCV, the present invention also relates to experimental infection of tamarins with the infectious GBV-B clone of the invention or with mutants of the infectious GBV clone to study indirectly the molecular properties of HCV or as a preliminary screen to identify 15 agents which have antiviral activity against HCV. For example, since the predicted internal ribosome entry site (IRES) structure in the 5'UTR of GBV-B is similar to that of HCV (Lemon 1997), the NS3 serine proteases of 20 GBV-B and HCV have been shown to share substrate specificity in vitro (Scarselli 1997), and the 3'UTRs of HCV (Yanagi 1999) and GBV-B (see Examples) have been 25 shown to be critical for viral infectivity, mutagenesis of these regions in the GBV-B infectious clone may be undertaken to examine IRES function, NS3 serine protease activity or the role of the 3'UTR in viral infectivity 30 in vivo. Where such "mutations" are introduced into the GBV-B clone of the invention to create a "mutated" GBV-B sequence, the mutations include, but are not limited to, point mutations, deletions and insertions. Of course, one of ordinary skill in the art would recognize that the size of the insertions would be limited by the 35 ability of the resultant nucleic acid sequence to be

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- ° properly packaged within the virion. Such mutations could be produced by techniques known to those of skill in the art such as site-directed mutagenesis, fusion PCR, and restriction digestion followed by religation.

5 Alternatively, given the significant structural homology that exists between the genomes of GBV and HCV, the infectious GBV-B clone may be used to screen for inhibitors of IRES function or viral enzyme activity (for example, NS3 helicase, NS3 protease, NS2-
10 NS3 protease or NS5B RNA polymerase activity). Such inhibitors may be useful as antiviral agents to HCV since viral enzyme activity and IRES function are known to be critical for HCV replication.

15 The effect of such inhibitors on the IRES function or viral activity of the GBV-B encoded by the infectious sequence of the invention may be measured by assays known to those of skill in the art to measure directly or indirectly viral replication or viral
20 pathogenicity. Such assays include, but are not limited to, the measurement of virus titer in serum or liver of an infected tamarin by PCR or the measurement of GBV-B viral protein expression in liver cells of an infected tamarin by immunofluorescence or Western blot. Of course, it is understood that a comparison of results obtained for control tamariins (treated only with infectious nucleic acid sequence) with those obtained
25 for treated tamariins (nucleic acid sequence and
30 antiviral agent) would indicate, the degree, if any, of antiviral activity of the candidate antiviral agent. Of course, one of ordinary skill in the art would readily understand that the tamariins can be treated with the
35 candidate antiviral agent either before or after

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- ° exposure to the infectious nucleic acid sequence of the present invention.

In yet another embodiment, the invention relates to "chimeric nucleic acid sequences" which consist of portions of the infectious nucleic acid sequence of GBV-B and portions of nucleic acid sequences of viruses which are related to GBV-B such as HCV, GBV-C and other members of the Flaviviridae family which do not infect tamarins. In a preferred embodiment, chimeric nucleic acid sequences consist of portions of the infectious nucleic acid sequence of GBV-B and portions of nucleic acid sequences of hepatitis C viruses (HCV) of various genotypes or subtypes; preferably portions of nucleic acid sequence of infectious HCV clones of genotypes 1a (ATCC accession number PTA-157; Figures 6A-6F), 1b (ATCC accession number 209596; Figures 7A-7F) or 2a (ATCC accession number PTA-153; SEQ ID NO: 4). The nucleic acid sequences taken from GBV-B and HCV can be open-reading frame sequences, and/or sequences from the 5'UTR and/or 3'UTR. The gene borders of the HCV genome, including nucleotide and amino acid locations, have been determined, for example, as depicted in Houghton, M. (1996), and the putative gene borders of the GBV-B are shown in Table 1.

Of course, it is understood that the production of GBV-B/HCV chimeras could include insertion of specific genes or regions of the infectious GBV-B clone into an HCV "genomic backbone" (where the HCV genomic backbone is preferably an infectious nucleic acid sequence of HCV genotypes 1a, 1b or 2a described above) or alternatively, could include insertion of

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° specific genes (or portions thereof) or regions of an HCV genome into the GBV-B infectious clone of the invention. Of course, where HCV genes or regions are to be inserted into the GBV-B infectious clone, it is to be understood that the inserted HCV sequences may be unmodified or may be mutated in order to examine the effect of the mutation(s) on the function of the inserted HCV gene or region in the chimeric GBV-B-HCV virus.

10 Such chimeras can readily be produced by methods known to those of ordinary skill in the art.

In one embodiment, GBV-B/HCV chimeras may be made in which 5' or 3' UTR sequences of the GBV-B infectious clone are replaced with the corresponding sequence from an HCV clone. For example, chimeras may be constructed in which the IRES sequence of the infectious GBV-B clone is replaced by the IRES sequence of HCV. Such chimeras can be used in identifying inhibitors of IRES activity which would be useful as antiviral agents, or could be used to examine HCV IRES function in vivo. Alternatively, mutations could be introduced into the HCV IRES contained in the GBV-B clone in order to examine the effect of the mutation(s) on IRES function in vivo.

30 Alternatively, GBV-B/HCV chimeras may be made in which the 3'UTR sequence of GBV-B is replaced by the 3'UTR sequence of HCV. As the 3' terminal stem-loop structure is believed to be important for initiation of RNA replication and has been shown to be critical for infectivity of HCV in vivo, such chimeras may be used for more detailed analysis of the function of the 3' UTR

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- o sequence of HCV in vivo and for the testing of candidate antiviral agents.

In another embodiment, GBV-B/HCV chimeras may be constructed in which the structural or non-structural regions of GBV-B are replaced by corresponding regions of HCV. Such chimeras would be useful in identifying whether the inability of HCV to infect tamarins is due to the inability of HCV's structural region to bind the receptor necessary for infection of tamarins or to the absence of sequences in HCV's nonstructural regions which are necessary for replication in tamarins. For example, the ability to infect tamarins with GBV-B/HCV chimeras in which the non-structural region of GBV-B is replaced by the non-structural region of HCV would indicate that the structural genes of GBV-B are necessary for viral infection in tamarins, and that the inability of HCV to infect tamarins is likely due to its lack of receptors for HCV.

Alternatively, the ability to infect tamarins with GBV-B/HCV chimeras in which the structural region of GBV-B is replaced by the structural region of HCV would indicate that the non-structural genes of GBV-B are critical for viral infection in tamarins, and that the inability of HCV to infect tamarins is likely due to HCV's lack of nonstructural sequences which are necessary for replication in tamarins.

Of course, GBV-B-HCV chimeras may be constructed in which only a portion of the non-structural or structural regions of GBV-B are replaced by the corresponding portions of HCV sequences. For example, a chimera in which only one or two of the three structural genes (C, E1 and E2) of GBV-B are replaced by

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° the corresponding HCV structural genes may be made. In one embodiment, nucleic acid sequences comprising the E1 and E2 genes of GBV-B may be replaced by the sequences comprising the HCV E1 and E2 genes. In another 5 embodiment, nucleic acid sequence comprising either the E1 or E2 gene of GBV-B is replaced by sequence encoding either the HCV E1 or E2 gene.

Alternatively, only a fragment of a GBV-B structural gene in the infectious GBV clone may be 10 replaced with the corresponding HCV gene fragments. For example, the amino terminal of the GBV-B E1 gene may be replaced by the corresponding portion of an HCV E1 gene or an amino terminal portion of the GBV-B E2 gene may be 15 replaced by an amino terminal portion of HCV E2 gene tht containing the HVR1 region. As the structural genes of HCV are believed to be important for neutralization, chimeras containing an HCV structural gene(s) or fragment(s) thereof can be used to develop vaccines 20 against HCV.

In yet another embodiment, chimeras in which individual non-structural genes of GBV-B, such as NS3 RNA helicase, NS3 protease, or the NS5B RNA-dependent 25 RNA polymerase are replaced by the corresponding non-structural genes of HCV may be constructed. Such chimeras would, for example, be useful in identifying inhibitors of viral enzyme activity which would be useful as antiviral agents. Of course, it is understood 30 that in order to construct chimeras in which the polyprotein cleavage sites of the GBV-B remain intact, it may be desirable to replace only a fragment of a nonstructural gene of GBV-B with the corresponding HCV 35 gene fragment.

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- The present invention also relates to polypeptides encoded by the nucleic acid sequences of the invention or fragments thereof. In one embodiment, said polypeptide or polypeptides may be fully or partially purified from viruses produced by cells transfected with the nucleic acid sequences of the invention. In another embodiment, the polypeptide or polypeptides may be produced recombinantly from a fragment of the nucleic acid sequences of the invention.
5 In yet another embodiment, the polypeptides may be chemically synthesized.
10

The present invention further relates to the in vitro and in vivo production of GBV-B, mutated GBV-B or chimeric GBV-B/HCV viruses from the nucleic acid sequences of the invention.
15

In one embodiment, the sequences of the invention can be inserted into an expression vector that functions in eukaryotic cells. Eukaryotic expression vectors suitable for producing high efficiency gene transfer in vivo are well known to those of ordinary skill in the art and include, but are not limited to, plasmids, vaccinia viruses, retroviruses, adenoviruses
20 and adeno-associated viruses.
25

In another embodiment, the sequences contained in the recombinant expression vector can be transcribed in vitro by methods known to those of ordinary skill in the art in order to produce RNA transcripts which encode the GBV-B of the invention. The GBV-B of the invention may then be produced by transfecting cells by methods known to those of ordinary skill in the art with either the in vitro transcription mixture containing the RNA
30
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- 15 -

- transcripts or with the recombinant expression vectors containing the nucleic acid sequences described herein.

In assaying the ability of the mutated GBV-B sequences or of the chimeric sequences of the invention to infect tamarins, the virulence phenotype of the virus produced by transfection of tamarins with the sequences of the invention can be monitored by methods known in the art such as measurement of liver enzyme levels (alanine aminotransferase (ALT) or isocitrate dehydrogenase (ICD)) or by histopathology of liver biopsies.

The present invention also relates to the use of the infectious GBV-B sequence, the mutated GBV-B nucleic acid sequences or the chimeric sequences of the invention to identify cell lines capable of supporting the replication of GBV-B or the chimeras of the invention.

Transfection of tissue culture cells with the nucleic acid sequences of the invention may be done by methods of transfection known in the art such as electroporation, precipitation with DEAE-Dextran or calcium phosphate, or incorporation into liposomes.

In one such embodiment, the method comprises the growing of animal cells in vitro and transfecting the cells with the nucleic acid of the invention, then determining if the cells show indicia of GBV-B or HCV infection. Such indicia include the detection of viral antigens in the cell, for example, by immunofluorescence procedures well known in the art; the detection of viral polypeptides by Western blotting using antibodies specific therefor; and the detection of newly transcribed viral RNA within the cells via methods such

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- o as RT-PCR. The presence of live, infectious virus particles following such tests may also be shown by injection of cell culture medium or cell lysates into healthy, susceptible animals, with subsequent exhibition of the signs and symptoms of GBV-B infection.

5 Suitable cells or cell lines for culturing GBV-B or the chimeric GBV-B-HCV include, but are not limited to, lymphocyte and hepatocyte cell lines known in the art.

10 Alternatively, primary hepatocytes can be cultured, and then infected; or, the hepatocyte cultures could be derived from the livers of infected tamarins. In addition, various immortalization methods known to those of ordinary skill in the art can be used to obtain cell-lines derived from hepatocyte cultures. For example, primary hepatocyte cultures may be fused to a variety of cells to maintain stability.

15 20 The invention also provides that the nucleic acid sequences and viruses of the invention be supplied in the form of a kit, alone or in the form of a pharmaceutical composition.

25 All scientific publication and/or patents cited herein are specifically incorporated by reference. The following examples illustrate various aspects of the invention but are in no way intended to limit the scope thereof.

30 EXAMPLES

Materials and Methods

Source of GB virus B

Two tamarin pools VR-806, (American Type Culture Collection) and H205, were used for experimental

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- ° transmission of the GB virus agents to tamarins species *Saguinus mystax* and *Saguinus oedipus*.

Amplification, cloning and sequence analysis of GBV-B
5 Viral RNA was extracted from aliquots of the
GB 2/94 serum pool or CT 11/91 liver homogenate with the
TRIzol system (GIBCO/BRL). Primers used in cDNA
synthesis and PCR amplification were based on the
genomic sequence of GBV-B published by Simons et al
10 (Simons 1995) shown in SEQ ID NO:3. Long RT-PCR was
performed using Superscript II reverse transcriptase
(GIBCO/BRL) and the Advantage cDNA polymerase mix
(Clontech) as described previously (Tellier 1996). Four
15 subgenomic regions of GBV-B covering the entire
published sequence (Simons 1995) were amplified from
serum and the PCR products were purified and cloned into
pGEM-9zf(-) (Promega) or pCR2.1 vector (Invitrogen)
using standard procedures.

20 The 5' terminus of GBV-B was amplified from
serum by using the rapid amplification of cDNA ends
(RACE) with dC or dA tailing (GIBCO/BRL) and GBV-B
specific antisense primers. Two different approaches
25 were used to determine the 3' terminal sequence of GBV-
B. In one approach, GBV-B RNA extracted from serum was
circularized with T4 RNA ligase (Promega) and the 5'-to-
3'-end-ligated viral RNA was amplified in RT-PCR using
specific GBV-B primers. In the second approach, the 5'
30 end of the negative strand GBV-B RNA extracted from the
liver homogenate was amplified using the 5' RACE with dc
tailing and GBV-B specific sense primers. The PCR
products were cloned directly into pCR2.1-TOPO by using
the TOPO TA Cloning Kit (Invitrogen).
35

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o The consensus sequence of GBV-B was determined by direct sequencing of PCR products (nucleotides 1-9078 and nucleotides 9130-9359) and by sequence analysis of the clones (nucleotides 1-7135 and nucleotides 7151-9399). Nucleotide positions correspond to those of the infectious clone (pGBB). Analyses of genomic sequences were performed with GeneWorks (Oxford Molecular Group) (Bukh 1995). To determine whether the GenBank data base contained sequences with homology to the GBV-B 3' UTR sequence identified in the present invention, a "Blast" search was performed. The predicted secondary structure of the GBV-B and HCV 3' UTR sequences were determined by the program "mfold" (Genetics Computer Group).

15 Construction of consensus cDNA clones of GBV-B

First, clone pGBB5-1, a consensus clone of GBV-B 2/94 containing the 3' terminus of GBV-B as published by Simons *et al* was constructed (Simons 1995a). The core sequence of the T7 promoter, a 5' guanosine residue and the sequence of GBV-B (9139 nucleotides) were cloned into pGEM-9zf(-) vector using *NotI/SacI* sites. A *BamHI* site was included at the GBV-B 3' terminus. Digested fragments containing the consensus sequence were purified from subclones and ligated using convenient sites. Next, a second consensus clone of GBV-B, clone pGBB, was constructed by inserting the additional 3' terminal sequence, amplified by PCR from one of the clones obtained by the RACE procedure described above, into pGBB5-1 using *XmaI* (at position 9114) and *BamHI* sites. A *XhoI* site was inserted following the GBV-B 3' terminus. DH5-alpha competent cells (GIBCO BRL) were transformed and selected on LB agar plates containing 100 µg/ml

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° ampicillin (SIGMA) and amplified in LB liquid cultures at 30°C for 18-20 hrs (Yanagi 1997). Each cDNA clone was re-transformed to select a single clone, and large-scale preparation of plasmid DNA was performed
5 with a QIAGEN plasmid Maxi kit as described previously (Yanagi 1997). Each clone was genetically stable since the digestion pattern was as expected following retransformation and the complete sequence was the expected one.

10 Intrahepatic transfection of tamarins with transcribed GBV-B RNA

In 100 µl reactions, RNA was transcribed *in vitro* with T7 RNA polymerase (Promega) from 10 µg of linearized template plasmid. The plasmid pGBB5-1 was linearized with *Bam*HI (Promega) and the plasmid pGBB was linearized with *Xho*I (Promega). The integrity of the RNA was checked by electrophoresis through agarose gel stained with ethidium bromide. Each transcription mixture was diluted with 400 µl of ice-cold phosphate-buffered saline without calcium or magnesium (SIGMA) and then immediately frozen on dry ice and stored at -80°C. Within 24 hours of synthesis, two transcription mixtures were injected into each tamarin by percutaneous intrahepatic injection guided by ultrasound (Emerson, 1992; Yanagi 1998, 1999). If the tamarin did not become infected, the same transfection
15 was repeated once. All transfected animals were negative for GBV-A_{SM} as determined by the protocol described previously (Bukh 1997a).

Monitoring of experimental course in tamarins

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o Serum samples were collected weekly from the tamarins and monitored for liver enzyme levels [alanine aminotransferase (ALT), gamma-glutamyltranspeptidase (GGT), and isocitrate dehydrogenase (ICD)] by standard methods and for GBV-B RNA by a specific reverse transcriptase-polymerase chain reaction (RT-PCR) assay. Total RNA was extracted from 100 µl of serum using the TRIzol reagent. The RNA pellet was resuspended in 10 mM dithiothreitol (DTT) containing 5% (vol/vol) of RNasin (20-40 u/µl) (Promega). The RT-nested PCR was performed with primers from the 5' UTR of GBV-B (external primer pair: 5'-CCT AGC AGG GCG TGG GGG ATT TCC-3' and 5'-AGG TCT GCG TCC TTG GTA GTG ACC-3'; internal primer pair: 5'-GGA TTT CCC CTG CCC GTC TG-3' and 5'-CCC CGG TCT TCC CTA CAG TG-3'). The reverse transcription was performed with avian myeloblastosis virus reverse transcriptase (Promega) and the external anti-sense primer and nested PCR was performed with AmpliTaq DNA polymerase or AmpliTaq Gold DNA polymerase (Perkin Elmer) as described previously (Bukh 1998a). Specificity was confirmed by sequence analysis of selected DNA products. Each set of experiments included a positive control sample (a 10⁻⁶ dilution of GB 8/93, estimated titer 100 genome equivalent (GE)) and appropriate negative control samples. The genome equivalent (GE) titer of GBV-B in positive samples was determined by RT-nested PCR on 10-fold serial dilutions of the extracted RNA (Bukh 1998a). One GE was defined as the number of GBV-B genomes present in the highest dilution positive in RT-nested PCR. The sensitivity of this RT-nested PCR assay for GBV-B was equivalent to that of our RT-nested PCR assay for HCV (Bukh 1998b), for example, conserved NS3

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° primers which had the same sensitivity for GBV-B as the 5' UTR primers could detect HCV at optimal sensitivity in samples with known HCV genome titer. Testing for GBV-A and GBV-A variants was performed by RT-nested PCR assays as described previously (Bukh 1997a).
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The consensus sequence of the complete ORF was determined by direct sequencing of overlapping PCR products obtained by long RT-nested PCR on serum from one of the tamarins infected with RNA transcripts as 10 previously described (Yanagi 1997).

Example 1

Transmission of GB Agent in Tamarins

To generate virus pools of the GB agent, 15 tamarins were inoculated intravenously with pooled sera of the eleventh tamarin passage of this agent (Fig. 1). Acute phase sera from a *S. mystax* tamarin which developed hepatitis were pooled (GB 8/93) and inoculated 20 into additional *S. mystax* tamarins to generate a second pool of acute phase serum (GB 2/94). Both serum pools contained approximately 10^8 GE/ml of GBV-B and GBV-A. A 10% liver homogenate (CT 11/91) was prepared from a *S. oedipus* tamarin which developed hepatitis following 25 inoculation with the twelfth passage of the GB agent. The titer of GBV-B in the liver homogenate was approximately 10^7 GE/ml. The GB 2/94 serum and CT 11/91 liver samples were used as GBV-B cloning sources in the 30 present study.

Inoculation of eight *S. mystax* tamarins with 35 ten-fold serial dilutions of the GB 2/94 pool demonstrated that its infectivity titer of GBV-B was 10^8 tamarin 50% infectious doses (TID₅₀) (Fig. 2). The five

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° GBV-B infected tamarins all developed acute resolving hepatitis characterized by early appearance of viremia (weeks 1 or 2 p.i.), peak viral titers of 10^7 - 10^8 GE/ml and clearance of viremia after 9-16 weeks (Fig. 2). Two
5 of these tamarins (*S. mystax* 769 and 777) were infected only with GBV-B and were negative for GBV-A and GBV-A_{SM}, whereas the other three tamarins were infected with both GBV-B and GBV-A_{SM}. A *S. mystax* tamarin inoculated with the liver homogenate also developed acute resolving
10 hepatitis with peak GBV-B titers of 10^7 GE/ml and clearance of viremia after 11 weeks. Likewise, four *S. mystax* tamarins inoculated with dilutions of the GB 8/93 pool developed acute resolving hepatitis with clearance
15 of the GBV-B virus after 11-26 weeks. Thus, GBV-B infection in *S. mystax* tamarins is characterized by acute hepatitis, early appearance of viremia, high peak viral titers and viral clearance.

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Example 2

Novel 3' Terminal Sequence of GBV-B

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The consensus sequence of the complete 5' UTR of GBV-B (nucleotides 1-445) was deduced from 13 clones containing nucleotides 1-283 and 3 clones containing nucleotides 31-445. In addition, the entire 5' UTR sequence was determined by direct sequencing of the amplicons. The sequences of the various clones were highly conserved and the consensus 5' UTR sequence of GBV-B from this pool was identical to that of the previously published sequence for GBV-B (Simons 1995a). It is noteworthy that 13 of 15 clones analyzed from the rapid amplification of cDNA ends (RACE) procedure contained the published GBV-B 5' terminus (A residue)

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- ° and that the same 5' terminus was obtained whether the 5' RACE was performed with dC or dA tailing.

The consensus sequence of the ORF (nucleotides 446-9037) was determined by direct sequencing of PCR products obtained using long RT-PCR (Yanagi 1997). In addition, 3 clones containing nts. 446-7135 (one of these clones had a deletion of nts. 3036-3636), 2 clones containing nts. 2019-3373, 5 clones containing nts. 7151-8261 and 7 clones containing nts. 7521-9037 were analyzed. The sequences of GBV-B clones in this pool were very homogeneous. Evidence of micro-heterogeneity was found at only 70 (0.8%) nucleotide and 36 (1.3%) amino acid positions, scattered throughout the ORF. The proportion of amino acid positions with heterogeneity ranged from 0.5-3.2% in different putative gene regions (lowest in NS3 and NS5B; highest in E2 and NS2). The GBV-B ORF sequence differed from the published sequence of GBV-B (Simons 1995) at 34 (0.4%) nucleotide and 12 (0.4%) deduced amino acid positions, respectively (Table 1).

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Table 1

Nucleotide and amino acid differences among GBV-B (Simons 1995a), the consensus sequence of GBV-B recovered from a virus pool used as the cloning source (GBV-B, 2/94) and the infectious clone of GBV-B (pGBB).

5	Genomic Region*	Position nt [aa]	Nucleotide			Amino Acid		
			GBV-B	GBV-B 2/94	pGBB	GBV-B	GBV-B 2/94	pGBB
	5' UTR (1-445)							
C (446-913)								
E1 (914-1489)	1030	C	T	T				
E2 (1490-2641)	1498	T	C (t)	C				
	1628 [395]	G	A (g)	A		V	I (V)	I
	2552 [703]	G	A (g)	A		D	N (D)	N
	2562,2563	C,A	A,C	A,C		P	H	H
	[706]							
	2566	T	T	T				
	2625 [727]	C	T	T		A	V	V
NS2 (2642-3385)	2647	C	T (c)	T				
	2816 [791]	C	T	T		L	F	F
	2855 [804]	A	G	G		T	A	A
	3235	A	G	G				
NS3 (3386-5125)	3475**	C	C (t)	T				
	3760	C	T (c)	T				
	4114	C	T	T				
	4117	C	A	A				
	4177	T	C	C				
	4615	C	T	T				
NS4A (5126-5290)								
NS4B (5291-6034)	5329	C	T	T				
	5332	T	C	C				
	5350	A	C	C				
	5455	C	T (c)	T				
NS5A (6035-7267)	6413	T	A (t)	A		L	M (L)	M
	[1990]							
	6577	G	T	T				
	6690	T	C (t)	C		I	T(I)	T
	[2082]							
	6965	T	C (t)	C		S	P (S)	P
	[2174]							
	7015	A	G (a)	G				
	7128	G	A	A		G	E	E
	[2228]							
	7138**	A	A	G				
	7142	A	G	G		T	A	A
	[2233]							
NSSB (7268-9037)	7282	T	C (t)	C				
	7849	C	A	A				
	7852	C	T	T				
	8942	G	A (g)	A		V	I (V)	I
	[2981]							
	8971	T	C	C				
	9026	C	T (c)	T				
3' UTR (9038-9399)	9067	T	C	C				
	Poly(U)	27 nts	11-23 nts	23 nts				
	9134	Deletion	C	C				
	9141-9399	ND	259 nts	259 nts				

*Nucleotide positions corresponding to pGBB. Putative gene borders defined as suggested by homology with HCV (Muerhoff 1995). No homology was observed at the NS2-NS3 junction.

**Positions that differ between the cloning source (GBV-B 2/94) and the infectious clone of GBV-B (pGBB). The change introduced into pGBB at position 7138 introduced an artificial SalI site. nd: Not determined. Nucleotides and amino acids shown in parenthesis were found as a minor species in the cloning source (GBV-B, 2/94)

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o The sequence for the 3' UTR is shown in Figure 3. Additional 3' UTR sequence was initially identified by performing RT-PCR across 5'-to-3'-end-ligated viral RNA extracted from serum. In all 4 clones with GBV-B sequences, the 5' UTR was truncated compared to the published sequence (simon 1995a). However, whereas one clone (29c) had the exact 3' terminus previously published by Simons et al. (Simons 1995a), the three other clones (29a, 29b, 29d) had 150 additional terminal nucleotides. Compared with the published sequence, all four clones had a single nucleotide insertion (C residue) at position 9134. Next, RACE using dC-tailing only was performed on the 5' end of the negative-strand RNA extracted from the liver homogenate. All 11 clones analyzed had additional sequences at the 3' terminus. Compared with the published GBV-B sequence, two clones (gb6, gb23) had 259 additional nucleotides, 8 clones (gb9, gb19, gb20, gb21, gb24, gb25, gb30, gb35) had 236 additional nucleotides and 1 clone (gb8) had 232 additional nucleotides. Moreover, all of these clones had the insertion at position 9134. The 3' UTR sequences among the various clones were highly conserved (Fig. 3). To demonstrate that the terminal 22 nucleotides found only in clones gb6 and gb23 existed in circulating viruses, RT-nested PCR was performed on 10-fold serially diluted RNA extracted from the serum pool GB 2/94 using an RT and external antisense primer deduced from this sequence. GBV-B RNA was detected at a dilution of 10^{-7} and the sequence of the amplicon was identical to the sequence recovered from the liver homogenate. Thus, the 3' UTR of GBV-B consists of a short sequence of 30 nucleotides followed by a 11-24

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- ° nucleotide-long poly (U) tract (single C residues were observed in GBV-B from the liver homogenate) and a 3' terminal sequence of at least 309 nucleotides. The new GBV-B 3' UTR sequence did not have significant homology 5 to any of the sequences deposited in the GenBank database. A prediction of the secondary structure of the 3' UTR sequence is shown in Figure 4. The most notable feature of the secondary structure is a highly stable stem-loop structure at the very 3' end consisting 10 of 47 nucleotides.

Example 3

The pGBB Clone of GBV-B is Infectious in vivo

15 The infectivity of RNA transcripts from the consensus clone pGBB5-1 which encompassed only the published GBV-B sequence (Simons 1995) was first tested. Within the GBV-B sequence there were no deduced amino acid differences and only 2 nucleotide differences (at 20 nucleotide positions 3475 and 7138) between the consensus sequence of the cloning source (GBV-B 2/94) and the sequence of pGBB5-1 clone. In addition, the 3' UTR of pGBB5-1 had a deletion at nucleotide position 25 9134 and was missing the 3' terminal 259 nucleotides (Fig. 3). Prior to transcription, the pGBB5-1 clone was linearized at the *BamHI* site with digestion at the exact GBV-B 3' terminus. The RNA transcripts from pGBB5-1 were injected into the liver of two tamarins (*S. mystax* 30 797 and 815). GBV-B RNA was not detected in weekly serum samples collected during 17 weeks of follow-up. As the susceptibility of these two tamarins to GBV-B was subsequently demonstrated by experimental infection 35 using a GBV-B virus pool, the consensus clone pGBB5-1

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- ° which lacks the 3' terminal sequence of GBV-B is thus not infectious *in vivo*.

Next, the infectivity of RNA transcripts from the full-length consensus GBV-B cDNA clone pGBB was tested. The pGBB clone was identical to the pGBB5-1 clone except in the 3' UTR. Thus, in addition to a 5' UTR of 445 nucleotides, an ORF of 8592 nucleotides encoding 2864 amino acids and a 3' UTR of 103 nucleotides, the pGBB clone also contains an additional 259 nucleotides in its 3' UTR. pGBB was linearized at the *Xho*I site which added an additional C residue at the 3' end of the transcribed GBV-B RNA. When RNA transcripts from the pGBB clone were injected into the liver of two tamarins (*S. mystax* 816 and 817), both tamarins became infected with GBV-B with viremia at week 1 p.i. and peak viral titers of 10^8 GE/ml (Fig. 5). The consensus sequence of PCR products of the complete ORF, amplified from serum obtained during week 2 p.i. from one tamarin (*S. mystax* 817), was identical to the sequence of pGBB, including at the two positions which differed from the consensus sequence of the cloning source and from the published sequence of GBV-B (Table 1). By performing RT-PCR as desired above, it was demonstrated that the very 3' terminal GBV-B sequence of pGBB existed in the circulating viruses in this tamarin. Within two weeks of the transfection both tamarins developed hepatitis with dramatically elevated liver enzyme levels (Fig. 5). Thus, the pGBB clone is infectious *in vivo* whereas the clone pGBB5-1 which lacks the last 259 nucleotides was not.

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◦ WHAT IS CLAIMED IS:

1. An isolated nucleic acid molecule which encodes GB virus-B, said molecule capable of expressing said virus when transfected into cells.

5 2. The nucleic acid molecule of claim 1, wherein said molecule encodes the amino acid sequence of SEQ ID NO:2.

10 3. The nucleic acid molecule of claim 2, wherein said molecule comprises the nucleic acid sequence of SEQ ID NO:1.

4. A DNA construct comprising a nucleic acid molecule according to claim 1.

15 5. A DNA construct comprising a nucleic acid molecule according to claim 3.

6. An RNA transcript of the DNA construct of claims 4 or 5.

20 7. A cell transfected with the DNA construct of claims 4 or 5.

8. A cell transfected with RNA transcripts of claim 6.

25 9. A GB virus-B polypeptide produced by the cell of claim 7.

10. A GB virus-B polypeptide produced by the cell of claim 8.

30 11. A GB virus-B produced by the cell of claim 7.

12. A GB virus-B produced by the cell of claim 8.

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- o 13. A GB virus-B whose genome comprises a nucleic acid molecule according to claim 1.
14. A GB virus-B whose genome comprises a nucleic acid molecule according to claim 3.
- 5 15. A method for producing a GB virus-B comprising transfecting a host cell with the DNA construct of claims 4 or 5.
- 10 16. A method for producing a GB virus-B comprising transfecting a host cell with the RNA transcript of claim 6.
- 15 17. A composition comprising a nucleic acid molecule of claim 1 suspended in a suitable amount of a pharmaceutically acceptable diluent or excipient.
18. A composition comprising a nucleic acid molecule of claim 3 suspended in a suitable amount of a pharmaceutically acceptable diluent or excipient.
- 20 19. A nucleic acid molecule comprising a chimeric virus genome, said genome being a GB virus-B genome according to claim 1 in which a 3' or 5' UTR sequence of the genome is replaced by a corresponding region of the 3' or 5' UTR sequence of a hepatitis C virus genome.
- 30 20. The nucleic acid molecule of claim 19, wherein a 3' UTR sequence of the genome of a GB virus-B is replaced by a corresponding 3' UTR sequence of a hepatitis C virus genome.
21. The nucleic acid molecule of claim 20, wherein the 3' UTR sequence is the 3' UTR terminal stem loop sequence.

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o 22. The nucleic acid molecule of claim 19,
wherein a 5' UTR sequence of the genome of a GB virus-B
has been replaced by a corresponding 5' UTR sequence of
a hepatitis C virus genome.

5 23. The nucleic acid molecule of claim 22,
wherein the 5' UTR sequence is the IRES sequence.

10 24. A nucleic acid molecule comprising a
chimeric virus genome, said genome being a GB virus-B
genome according to claim 1 in which the non-structural
region of the genome of a GB virus-B has been replaced
by the non-structural region of a hepatitis C virus
genome.

15 25. The nucleic acid molecule of claim 24,
wherein at least one gene from the non-structural region
of the genome of a GB virus-B has been replaced by the
corresponding gene from the non-structural region of a
hepatitis C virus genome.

20 26. The nucleic acid molecule of claim 25,
wherein the gene from the non-structural region is
selected from the group consisting of NS3 protease, NS3
RNA helicase, or NS5B RNA polymerase.

25 27. A nucleic acid molecule comprising a
chimeric virus genome, said genome being a GB virus-B
genome according to claim 1 in which the structural
region of the genome of a GB virus-B has been replaced
by the structural region of a hepatitis C virus genome.

30 28. The nucleic acid molecule of claim 27,
wherein at least one gene from the structural region of
the genome of a GB virus-B has been replaced by the

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- o corresponding gene from the structural region of a hepatitis C virus genome.

29. The nucleic acid molecule of claim 28, wherein the gene from the structural region is selected
5 from the group consisting of E1, E2 or C.

30. The nucleic acid molecule of claim 28, wherein the E1 and E2 genes from the structural region of the genome of a GB virus-B have been replaced by the
10 E1 and E2 genes of a hepatitis C virus genome.

31. The nucleic acid molecule of claim 28, wherein the E1 gene from the structural region of the genome of a GB virus-B has been replaced by the E1 gene
15 of a hepatitis C virus genome.

32. The nucleic acid molecule of claim 28, wherein the E2 gene from the structural regions of the genome of a GB virus-B has been replaced by the E2 gene
20 of a hepatitis C virus genome.

33. A DNA construct comprising the nucleic acid molecule of claims 19, 24 or 27.

34. An RNA transcript of the DNA construct of
25 claim 33.

35. A virus whose genome comprises a nucleic acid molecule according to claims 19, 24 or 27.

36. A nucleic acid molecule comprising a chimeric virus genome, said genome being a hepatitis C virus genome in which a 3' or 5' UTR sequence of the genome is replaced by a corresponding region of the 3' or 5' UTR sequence of a GB virus-B genome according to
30 claim 1.

35

- 36 -

o 37. A nucleic acid molecule comprising a
chimeric virus genome, said genome being a hepatitis C
virus genome in which the non-structural region of the
genome has been replaced by the non-structural region of
5 a GB virus-B genome according to claim 1.

10 38. A nucleic acid molecule comprising a
chimeric virus genome, said genome being a hepatitis C
virus genome in which the structural region of the
genome has been replaced by the structural region of a
GB virus-B genome according to claim 1.

39. A polypeptide encoded by the nucleic acid
molecule of claims 19, 24 or 27.

15 40. A polypeptide encoded by the nucleic acid
molecule of claims 36, 37 or 38.

20

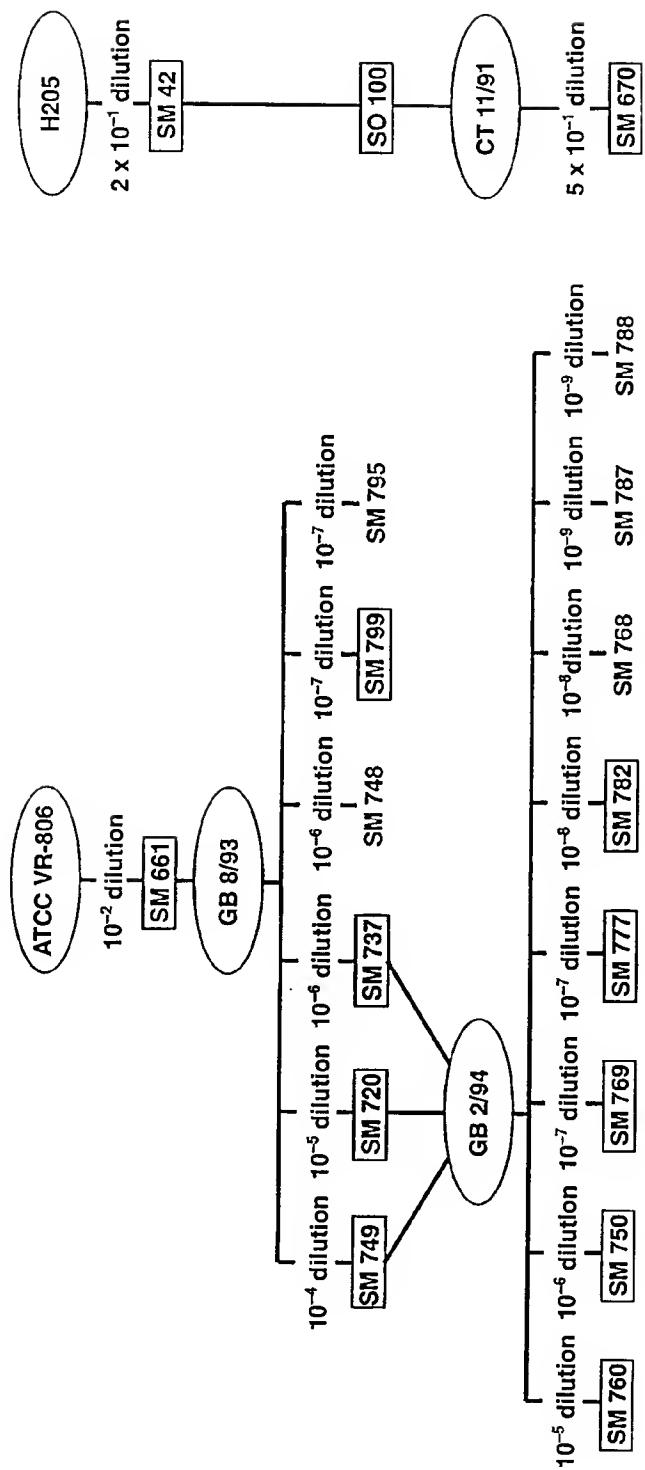
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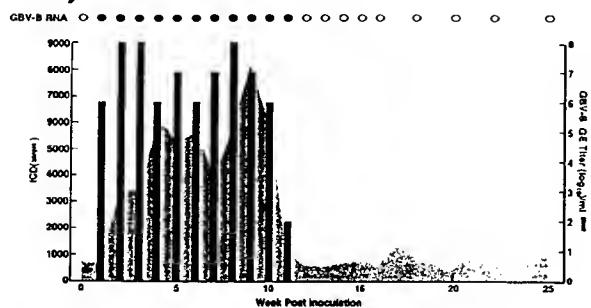
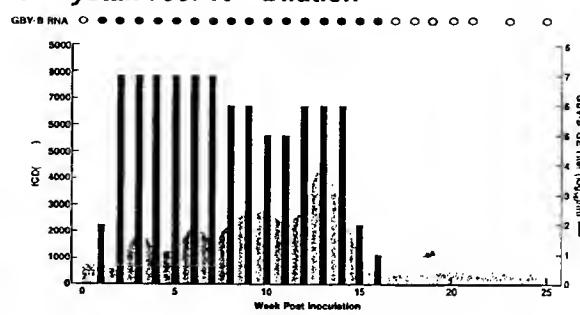
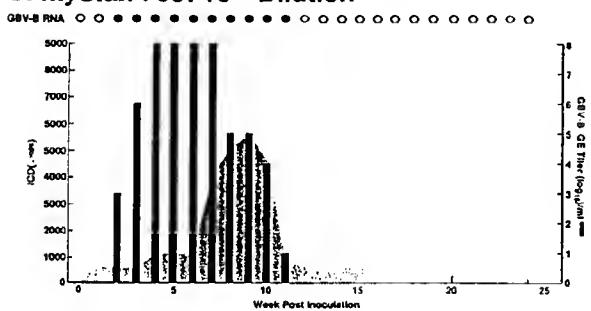
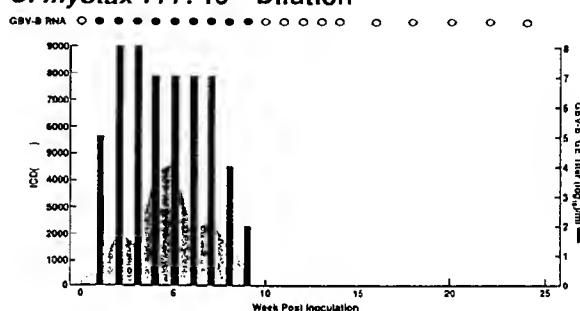
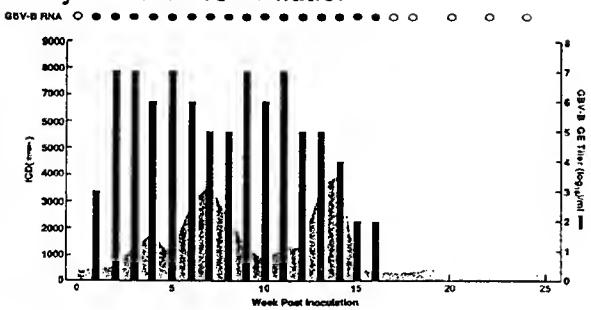
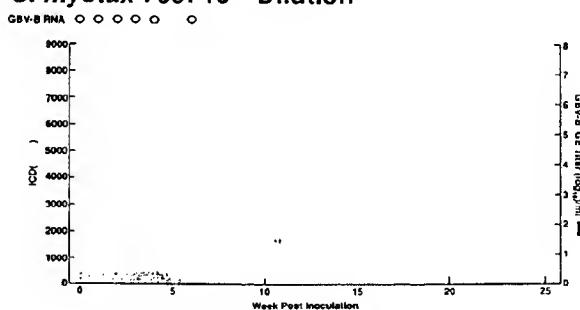
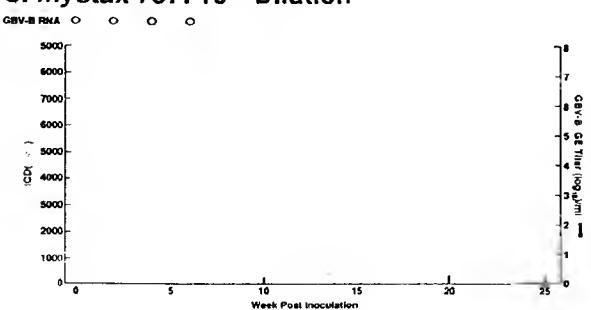
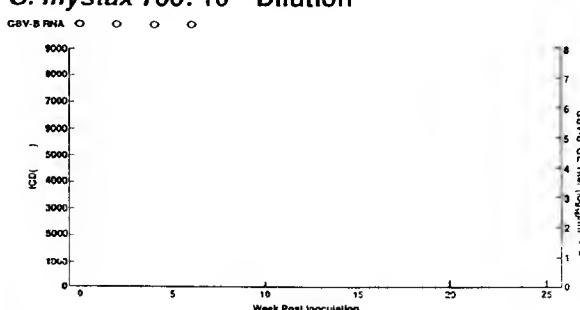
35

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FIG. 1



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FIG. 2***S. mystax* 760: 10^{-5} Dilution*****S. mystax* 750: 10^{-6} Dilution*****S. mystax* 769: 10^{-7} Dilution*****S. mystax* 777: 10^{-7} Dilution*****S. mystax* 782: 10^{-8} Dilution*****S. mystax* 768: 10^{-8} Dilution*****S. mystax* 787: 10^{-9} Dilution*****S. mystax* 788: 10^{-9} Dilution**

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FIG. 3

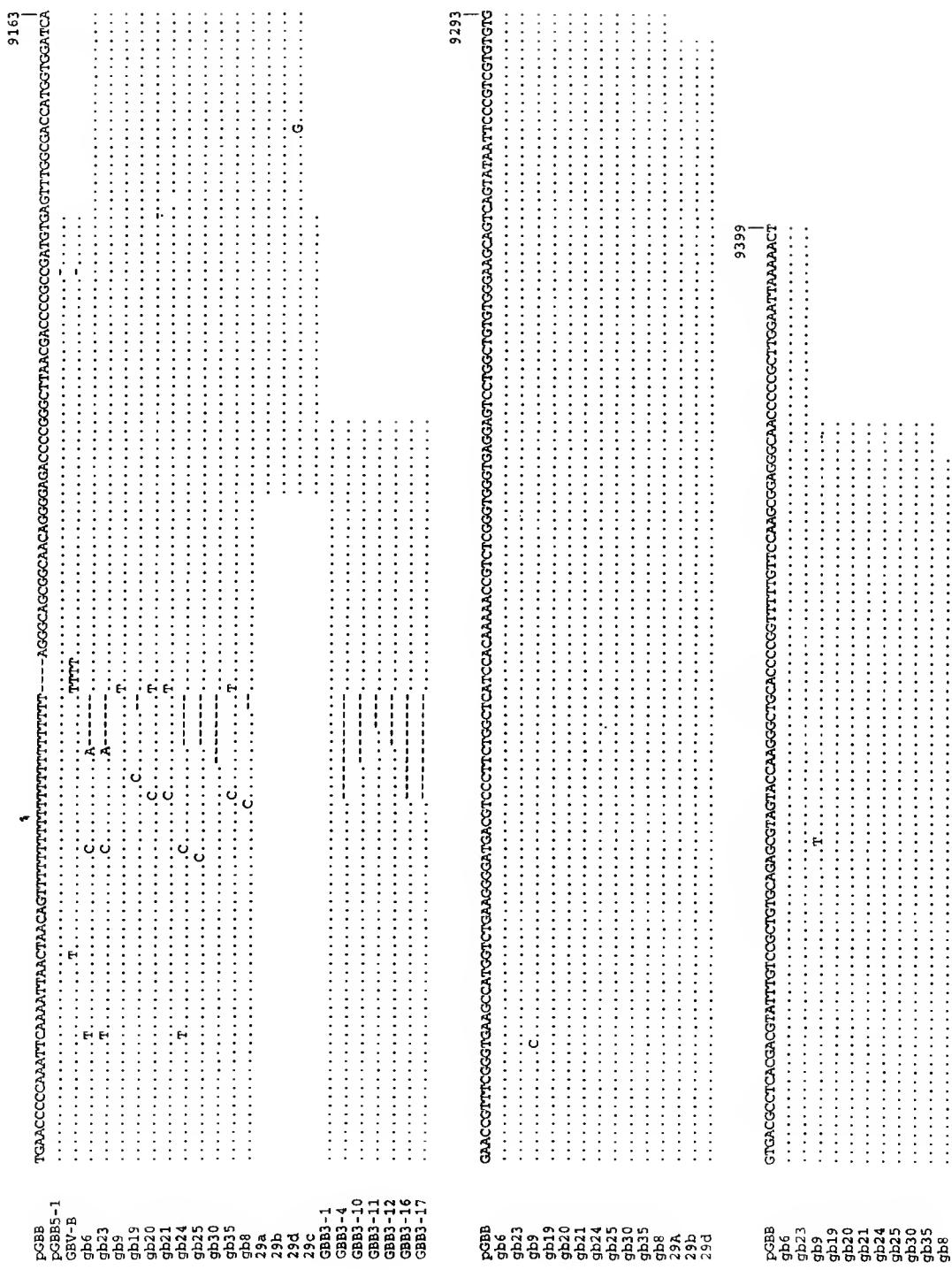
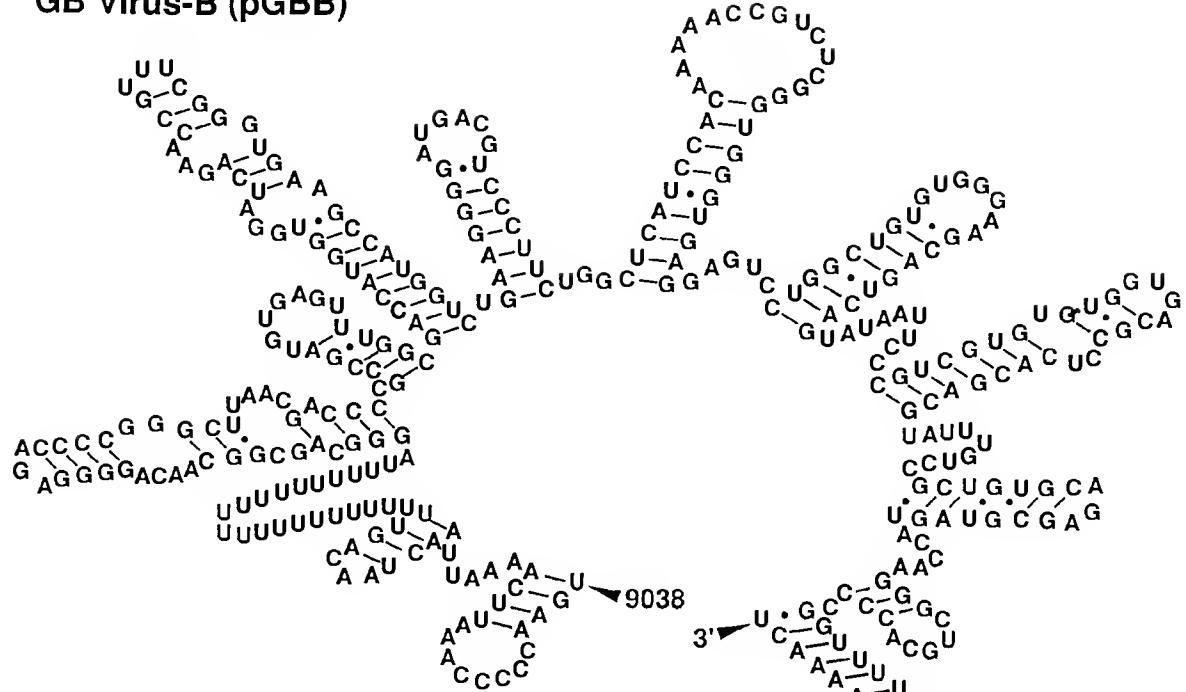
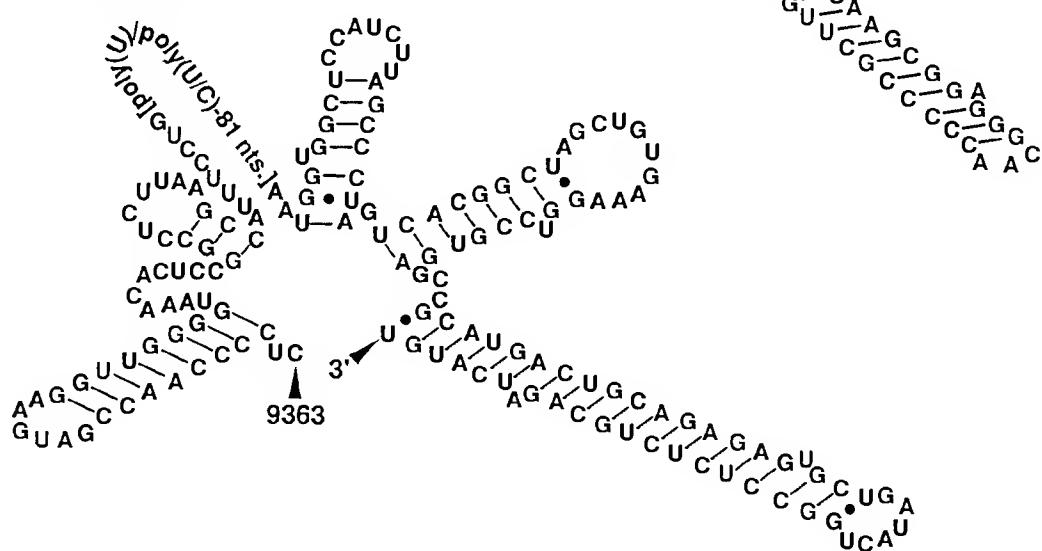


FIG. 4

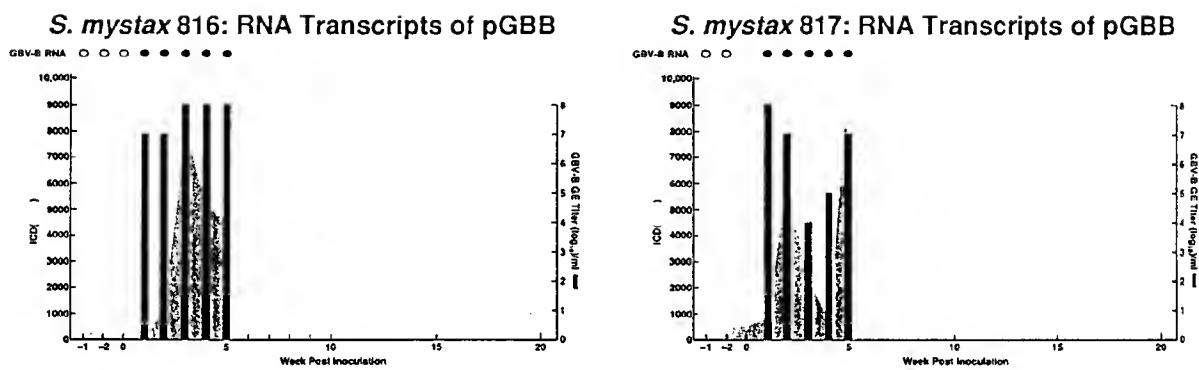
GB Virus-B (pGBB)



Hepatitis C Virus (pCV-H77C)



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FIG. 5

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GCCAGCCCCC	TGATGGGGGC	GACACTCCAC	CATGAATCAC	TCGGCTGTGA	50
GGAACTACTG	TCTTCAOGCA	GAAAGCGTCT	AACCATGGCG	TTAGTATGAG	100
TGTCGTCAG	CTTCCAGGAC	CCCCCTTCCC	GGGAGAGCCA	TAGTGGCTTG	150
CGGAACCGGT	GAGTACACCG	GAATTGOCAG	GAAGAACGGG	TCCTTCTTIG	200
GATAAAACCG	CTCAATGCGT	GGAGATTTCG	GGTGCACCC	GCAAGACTGC	250
TAGCCGAGTA	GTGTTGGTC	GOGAAAGGCC	TIGTGGTACT	GCCTGATAGG	300
GTGCTTGCAG	GTGCCCCGGG	AGGTCCTCGA	GAACGTGCAC	CATGAGCAOG	350
AATCCTAAAC	CTCAAAGAAA	AACCAAAACGT	AACACCAAAAC	GTGGCCACAA	400
GGAOGTCAAG	TTCCCCGGTG	GGGGTCAGAT	CGTTGGTGGG	GTTTACTTGT	450
TGCCGGCGAG	GGGCCCCAGA	TTGGGTGTC	GGGGAGAGAG	GAAGACTTCC	500
GAGGGTGC	AAACCTCGAGG	TAGACGTCAG	CCATATCCCCA	AGGCAACGTG	550
GGGGAGGGC	AGGACCTGGG	CTCAGCCCCG	GTACCCCTTG	GGGCTCTATG	600
GCAATGAGGG	TTGGGGGTGG	GGGGGATGGC	TGCTGTCCTC	GGGGGGCTCT	650
GGGGCTAGCT	GGGGCCCCAC	AGACCCCCCG	CGTAGGTGCG	GCAATTGGG	700
TAAGGTCAATC	GATACCCCTA	CGTGACGGCTT	GGGGGACCTC	ATGGGGTACA	750
TACCGCTCGT	GGGGGCCCCCT	CTTGGAGGGG	CTGGCCAGGGC	CCTGGGGCAT	800
GGGGTCAGGG	TTCTGGAAGA	GGGGGTGAAC	TATGCAACAG	GGAACCTTCC	850
TGGTTGCTCT	TCTCTATCT	TCCTTCGTC	CTCTCTCTCT	TGCTGACTTG	900
TGCCCGCTTC	AGCTTACCAA	GTGGCCAATT	CTCGGGGGCT	TTACCATGTC	950
ACCAATGATT	GGGCTAACTC	GAGTATTGIG	TACGGAGGGG	GGGATGCCAT	1000
CCTGCACACT	GGGGGGTGTG	TCCTTGGGT	GGGGGAGGGT	AACGCGTGA	1050
GGTGTGGGT	GGGGGTGACC	CCCCGGTGG	CCACCAAGGA	GGGCAAACTC	1100
CCCCACAACGC	AGCTTGCACG	TCAATATCGAT	CTGCTTGTG	GGAGGGCCAC	1150
CCCTCTGCTCG	GGGCTCTACG	TCGGGGACCT	GTGGGGGTCT	GTCTTCTTIG	1200
TIGGTCAACT	GTTAACCTTC	TCTCCCAAGG	CCACCTGGAC	GACGCAAGAC	1250
TGCAATTGGT	CTATCTATCC	GGGCGATATA	AGGGTCAATC	GCATGGCATG	1300
GGATATGATG	ATGAACCTGGT	GGGCTACGGC	AGCGTGGTG	GTAGCTCAGC	1350
TGCTCCGGAT	GGGCGATATA	ATCATGGACA	TGATGGCTGG	TGCTCACTGG	1400
GGAGTGGCTG	GGGGCATAGC	GTATTTCTCC	ATGGTGGGGA	ACTGGGGGAA	1450
GGTGGCTGGTA	GTGCTCTTC	TATTTGGCGG	GGTGGACGGG	GAAACCCACG	1500
TCACCGGGGG	AAATGGGGCC	GGGCGATATA	CTGGGGCTGT	TGGCTCTCTT	1550
ACACCAAGGG	GGGCGATATA	GGGCGATATA	GGGCGATATA	GGGCGATATA	1600
GCACATCAAT	AGCAACGGCT	TGAATTGCAA	TGAAAGCCCT	AAACACCGGCT	1650
GGITAGCAGG	GCTCTTCTAT	CAACACAAAT	TCAACTCTTC	AGGCTGTGCT	1700
GAGAGGTTGG	GGGGCTGGG	GGGGCTGGG	GGGGCTGGG	GGGGCTGGG	1750
TCCTATCAGT	TATGCGAAACG	GGGGCTGGG	GGGGCTGGG	GGGGCTGGG	1800
GGCACTAACCC	TCCAAGACCT	TGTTGGCTTG	TGTTGGCTTG	GGGGCTGGG	1850
GGGGGGTAT	ATGGCTTCAC	GGGGCTGGG	GGGGCTGGG	GGGGCTGGG	1900

FIG. 6A

H77C

10	20	30	40	50
1234567890	1234567890	1234567890	1234567890	1234567890
CAGGTGGGGC	GCGGCTAACCT	ACAGCTGGGG	TGCAAATGAT	AOGGATGTCT
TOGTCCTTAA	CAACACCAGG	CCAOOGCTGG	GCAATTGGTT	OGGTIGTGTACC
TGGATGAACT	CAACCTGGATT	CACCAAAGTG	TGCGGAGGCG	CCCGCTTGCT
CATCGGAGGG	GTGGGCAACA	ACAOCTTGCT	CTGCCCCACT	GATTGCTTCC
GCAAACATCC	GGAAGGCCACA	TACTCTGGT	CGGGCTCGGG	TGCCCTGGATT
ACACCCAGGT	GCATGGTGA	CTACCCGAT	AGGCTTTGGC	ACTAATCCTTG
TACCATCAAT	TACACCATAT	TCAAAGTCAG	GATGTAOGTG	GGAGGGGGTGT
AGCACAGGCT	GGAAGGCGGC	TGCAACTGGA	CGGGGGGGGA	AOGCTIGTGTAT
CTGGAAGACA	GGGACAGGTC	CGAGCTCAGC	CGGTTGCTTC	TGTCACACCAC
ACAGTGGCAG	GTGCTTGGT	GTGCTTTCAC	GACGCTGCGA	GCCTTGTGCGA
CGGGGCTCAT	CCAOCTCCAC	CAGAACATTG	TGGACGTGCA	GTACTTGTAC
GGGGTAGGGT	CAAGCATCGC	GTGCTGGGCC	ATTAAGTGGG	AGTACGTCGT
TCTCTGTTTC	CTTCTGCTTG	CAGACGGGGG	CGTCTGCTCC	TGCTTGTGGA
TGATGTTACT	CATAATCCAA	GGGGAGGGG	CTTGGAGAAA	CTCTGTAATA
CTCAATGCAG	CATCCCTGGC	CGGGACGGAC	GGTCTTGTGT	CCTTCCTCGT
GTGCTCTGTC	TTTGGGTTGT	ATCTGAAGGG	TAGGTCGGTG	CCCCGGAGCGG
TCTACGGGCT	CTACGGGATG	TGGGCTCTCC	TCCTGCTCT	GCTGGGGTTTG
CCTCAGGGGG	CATA CGCACT	GGACACGGAG	GTGGGGCGGT	CGTGTGGCGG
CGTGTGTCIT	GTGGGGTAA	TGGGCGCTGAC	TCTGTOGCCA	TATTACAAGC
GCTATATCAG	CTGGTGCATG	TGGTGGCTTC	AGTATTTCT	GACCAGAGTA
GAAGCGAAC	TGCACGTGIG	GGTTCGGGGG	CTCAACGTCC	GGGGGGGGCG
CGATGCGTC	ATCTTACTCA	TGIGTGTAGT	ACACCCGACC	CTGGTATTTG
ACATCACCAA	ACTACTCTTG	GCCATCTTCG	GACCCCTTTC	GATTCCTCAA
CCCAGTTGTC	TTAAAGTCCC	CTACTTGTGIG	CGGGTTCAAG	GCCTTCTCCG
GATCTGCGCG	CTAGGGGGGA	AGATAGGGG	AGGTCAATTAC	GTGCAAATGG
CCATCATCAA	GTGAGGGGG	CTTACTGGCA	CTTATGIGTA	TAACCATCTC
ACCCCTCTTC	GAGACTGGGC	GCACAAACGGC	CTGCGAGATC	TGGCGTGGC
TGTGGAACCA	GTGGTCTTCT	CGCGAATGGA	GACCAAGCTC	ATCAOGTGGG
GGGCAGATAC	CGGGCGGTGC	GGTGACATCA	TCAACGGCTT	GGGGCTCTCT
GGGGGTAGGG	GCCAGGAGAT	ACTGCTGGG	CCACCGGAOG	GAATGGTCTC
CAACGGGTGG	AGGTCTCTGG	CGGGCATCAC	GGCGTACGCC	CACCAAGCGA
GAGGGCTCT	AGGGIGTATA	ATCACCAAGCC	TGACTGGGGG	GGACAAAAAC
CAAGTGGAGG	GTGAGGTCCA	GATGGTGTCA	ACTGCTACCC	AAACCTTCT
GGCAAGTGT	ATCAATGGGG	TAIGCTGGAC	TCTGTACAC	GGGGGGGGAA
CGAGGACCAT	CGCATCACCC	AAGGGTCTTG	TCATOCAGAT	GTATACCAAT
GTGGACCAAG	ACCTTGTGGG	CTGGGGCGCT	CTCAAGGTT	CCCGCTCAIT
GACACCCGT	ACCTGCGGCT	OCTCGGACCT	TIAACCTGGTC	ACGAGGGCAG
CGATGTCAT	TCCCGTGGCG	CGGGGAGGTG	ATAGCAGGGG	TAGGCTGCTT
				3800

FIG. 6B

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H77C

10	20	30	40	50
1234567890	1234567890	1234567890	1234567890	1234567890
TGCCCCGGC	CCATTCTTA	CITGAAAGGC	TCTCTGGGGG	GTCGGCTGTT
GTGCCCCGGG	GGACACCGGG	TGGGCTTATT	CAGGGGGGGG	GTTGTCGAACC
GTGGAGCTGC	TAAAGCGGTG	GACTTTATCC	CITGTTGAGAA	CCTAGGGACA
ACCATGAGAT	CCCCGGTGTG	CAOGGACAAC	TCTCTCTCAC	CAGCAGTGCC
CCAGAGCTTC	CAGGTGGGGC	ACCTGCTATGC	TCCCACGGGC	AGCGGTAAGA
GCACCAAGGT	CCCGGCTGCG	TACGCAGGCC	AGGGCTACAA	GGTGTGGTG
CTCAACCCCT	CTGTTGCTGC	AACGCTGGGC	TTTGGTGCCT	ACATGTCAAA
GGCCCATGGG	GTGATCTTA	ATATCAGGAC	CGGGGTGAGA	ACAATTAA
CTGGCAGGCC	CATCAOGTAC	TOCACTTAAG	GCAAGTTCCT	TGCGGACGGC
GGGTGCTCAG	GAGGTGCTTA	TGACATAATA	ATTTGIGAOG	AGTGCACTC
CAOGGATGCC	ACATCCTATCT	TGGCCATOGG	CACTGTCCTT	GACCAAGCAG
AGACTGCGGG	GGCGAGACTG	GTGTTGCTCG	CCACTGCTAC	CCCTCGGGGC
TCGGTCACTG	TGTCCCATOC	TAACATCGAG	GAGGTTGCTC	TGTCACCCAC
CGGAGAGATC	COCTTTAACG	GCAAGGCTAT	CCCCCTCGAG	GTGATCAAAG
GGGGAAGACA	TCTCATCTTC	TGCCACTCAA	AGAAGAAGTG	CGACGAGCTC
GGCGCGAAGC	TGGTCGCATT	GGGCATCAAT	CCCCGTGGCT	ACTACOGGG
TCTTGACGTG	TCTGTCATCC	CGACCAGCGG	CGATGTTGTC	GTGTTGTCGA
CCGATGCTCT	CATGACTGGC	TTTACCGGGG	ACTTCGACTC	TGTGATAGAC
TGCAACACGT	GTGTCACTC	GACAGTGGAT	TTCAAGCTTG	ACCCCTACCTT
TACCAATTGAG	ACAACCACGC	TOCCCCAGGA	TGCTGTCCTC	AGGACTCAAC
GGCGGGGGAG	GAATGGCAGG	GGGAAGCCAG	GCATCTATAG	ATTTGIGGCA
GGGGGGGAAC	GGGGCTCCGG	CATGTTGAC	TGTCACGTC	TCTGTGAGTG
CTATGACGGG	GGCTGTCCTT	GGTATGAGCT	CAACGGGGGCC	GAGACTACAG
TTAGGCTACG	AGGGTACATG	AACACCCCGG	GGCTTCCCGT	GTGCCAGGAC
CATCTTGAAT	TTTGGGAGGG	CGCTTTAACG	GGCTCACTC	ATATAGATCC
CCACTTTITA	TCCCAGACAA	ACCAGAGTCG	GGAGAACTTT	OCTTAOCTGG
TAGCGTACCA	AGCCACCGTG	TGGCTTAGGG	CTCAAGCCCC	TOCCCCATCG
TGGGACCAGA	TGTTGAAAGTG	TTTGATCGC	CTAAACCCA	CCCTOCATGG
GCACACACCC	CTGCTATACA	GAATGGGGCG	TGTTCAAGAT	GAAGTCACCC
TGAOGCAACC	AATCACCAAA	TACATCATGA	CAATGATGTC	GGCGACCTG
GAGGTGCGTC	CGAGCAACCTG	GGTGTCTGTT	GGCGGGGTC	TGGCTGCTCT
GGCGCGTAT	TGCGTGTCAA	CAGGCTCGT	GGTCATAGTG	GGCAGGATCG
TCTTGTCGGG	GAAGGGGGCA	ATTATACCTG	ACAGGGAGGT	TCTCTACCG
GAGTTCGATG	AGATGGAAGA	GTGCTCTCAG	CACTTACCGT	ACATOGAGCA
AGGGATGATG	CTCGCTGAGC	AGTTCAAGCA	CAAGGGCCCTC	GGCTCTCTGC
AGACCGGGTC	CGCGCATGCA	GAGGTATICA	CCCGCTGCTG	CCAGACCAAC
TGGCAGAAC	TCGAGGTCTT	TTGGGCGAAG	CACATGTCGA	ATTTCATCAG
TGGGATACAA	TACTTGGGGG	GGCTGTCAAC	GCTGCTGGT	AACCCCGOCA
				5700

H77C

10	20	30	40	50
1234567890	1234567890	1234567890	1234567890	1234567890
TTGCTTCATT	GATGGCTTT	ACAGCTGCG	TCAACCAGCC	ACTAACCAC
GGCCAAACCC	TCCCTCTCAA	CATATTCGGG	GGGTGGGTGG	CTGCCCCAGCT
CGGGGCCCCC	GGTGCAGGCTA	CTGGCTTGT	GGGTGGCTGGC	CTAGCTGGG
CGGACATCGG	CAAGCTTCTGA	CTGGGGAGG	TOCTOGTGG	CATTCTTGCA
GGGTATGGGG	GGGGAGTGGC	GGGAGCTCTT	GTAGCATTCA	AGATCATGAG
CGGTGAGGGC	GGCTCCACGG	AGGAAGCTGG	CAATCTGCTG	GGGGGCGATOC
TCTCGOCTGG	AGGCGCTTGT	GTGGGGTGG	TCTGCGCAGC	AATACTGGCG
CGGCAOGTTG	GGGGGGGGGA	GGGGGGAGTG	CAATGGATGA	GGGGGCTTAAT
AGGCTTCGCC	TCGGGGGGGA	ACCATGTTTC	GGGGGCGCAC	TAAGTGGCGG
AGAGCGATGC	AGGCGCCCGC	GTCACTGCGA	TACTCAGCG	GGGGGGCTGTA
ACCCAGCTCC	TGAGGGGACT	GCATCAGTGG	ATAAGCTGG	AGTGTACCCAC
TCCATGCTCC	GGTCTCTGGC	TAAGGGACAT	CTGGGACTGG	ATATGCGAGG
TGCTGAGCGA	CITTAAGACC	GGGCTGAAAG	CCAAGCTCAT	GGGACAACTG
CCTGGGATTG	CCTTGTGTC	CTGGCGAGGC	GGGTATAGGG	GGGTGGCTGGG
AGGAGAOGGC	ATTATGCCACA	CTGGCTGCGA	CTGTCGGAGC	GAGATCACTG
GACATGTCAA	AAACGGGACG	ATGAGGATCG	GGGGGCTTAG	GGGGGGCGAGG
AACATGTGGA	GTGGGACGTT	GGGGGATTAAC	GGCTACACCA	GGGGGGGGCTG
TACTCCCCCTT	CCTGCGCGGA	ACTATAAGTT	GGGGCTGTTG	AGGGGTTCTG
CAGAGGAATA	GGGGGAGATA	AGGCGGGTGG	GGGACTTCGA	CTACGTATCG
GGTATGACTA	CTGACAATCT	TAAATGCCCG	TGGGAGATCC	CAATGGGGCGA
ATTTTTCACA	GAATTGGGAGG	GGGTCGGGCT	ACACAGTTT	GGGGGGGGCTT
GCAAGCCCTT	GCTGGGGGAG	GGGGTATCAT	TCAGAGTAGG	ACTCCACGGAG
TACCCGGTGG	GGTGGCAATT	ACCTTGGAG	GGGGGACCGG	AGGTGGGGGT
GTGACGTCC	ATGCTCACTG	ATCCCTGCCA	TATAACAGCA	GGGGGGGGCG
GGAGAAGGGT	GGGGAGAGGG	TCACCCCTT	CTATGGCCAG	CTGGGGGGCT
AGCCAGCTGT	GGGGCTCCATC	TCTCAAGGCA	ACTTGGCACCG	CCAACCATGA
CTCCCCCTGAC	GGGGAGCTCA	TAGAGGCTAA	CTGGGGTGG	AGGCAGGGAGA
TGGGGGGCAA	CATCACCAAGG	GTGAGTCAG	AGAACAAAGT	GGGTGATTCTG
GACTCCCTCG	ATCCGCTTGT	GGGGAGGGAG	GGGGGGGGGG	AGGTGCTGGT
ACCTGGAGAA	ATTCCTGGGA	AGTCTGGAG	ATTCGGGGGG	GGGGGGGGCG
TCTGGGCGCG	GGGGGACTAC	AAACCCCGGC	TAGTGGAGAC	GGGGAAAAG
CCTGACTAACG	AAACCACTGT	GGTGGCATGGC	GGGGGGCTAC	GGGGGGGGCG
GTCCCCCTCCT	GTGGCTCCGC	CTGGGAAAAAA	GGGTACGGTG	GGGGGGGGCG
AAATCAACCC	ATCTACTGCC	TTGGGGAGC	GGGGGGGGGG	GGGGGGGGCG
AGCTTCTCAA	CTTGGGGCAT	TAAGGGGGAC	AAATACGACAA	GGGGGGGGCG
GGGGGGGGCG	TCTGGGGTCCC	GGGGGGACTC	GGGGGGGGGG	GGGGGGGGCG
CCATGGGGCC	GGGGGGGGGG	GGGGGGGGGG	GGGGGGGGGG	GGGGGGGGCG
TCATGGTGA	GGGTGAGTAG	GGGGGGGAC	GGGGGGGGGG	GGGGGGGGCG

FIG. 6D

1921

H77C

10	20	30	40	50
1234567890	1234567890	1234567890	1234567890	1234567890
CTCAATGCT TATTCCTGG AAGGGCGACT CGTCACCCCG TGCGCTGGGG				7650
AAGAACAAAA ACTGCCCATC AAOGCACTGA GCAACTCGTT GCTAOGCCAT				7700
CACAATCTGG TGTATTCCAC CACTTCAAGC AGTGCTTGCC AAAGGCAGAA				7750
GAAAGTCACA TTTGACAGAC TGCAAGTTCT GGACAGCCAT TACCAGGAOG				7800
TGCTCAAGGA GGTCAAAGCA GCGGGGTCAA AAGTGAAGGC TAACCTGCCTA				7850
TCGGTAGAGG AAGCTTGCGAG CCTGAOGCCC CCACATTCAAG CCAAATCCAA				7900
GTTCGGCTAT GGGCAAAAG ACGTCGGTIG CCATOCAGA AAGGCGGTAG				7950
CCACACATCAA CTGGTGTGG AAAGACCTTC TGGAAAGACAG TGTAACACCA				8000
ATAGACACTA CCATCATGGC CAAGAACGAG GTTTTCTGGG TTCAAGCTGA				8050
GAAGGGGGGT CGTAAGCAG CTGGTCTCAT CGTGTTCOCCC GAACCTGGCG				8100
TGCGCGTGTG CGAGAAGATG GCGCTGTACG ACGTGGTTAG CAAGCTTCOCCC				8150
CTGGCGGTGA TGGGAAGCTC CTACGGATTG CAATACTCAC CAGGACAGCG				8200
GGTGAATTTC CTGGTGCAAG CGTGGAAAGTC CAAGAAGACC CGATGGGT				8250
TCTCGTATGA TACCCGCTGT TTTGACTCCA CAGTCACTGA GAGGGACATC				8300
CGTAOGGAGG AGCCAATTIA CCAATGTTGT GAACCTGGACC CCCAAGCCG				8350
CGTGGCCATC AAGTCCCTCA CTGAGAOGCT TTATGTTGGG GCGCTCTTA				8400
CCAATTCAAG GGGGAAAAC TGCGGCTTAC GCAGGTGGCG CGCGAGCGC				8450
GTACTGACAA CTAGCTGTG TAACACCCCTC ACTTGCTACA TCAAGGCCCC				8500
GGCAGCTGT CGAGCGCGAG GGCTCAGGA CTGCACCATG CTGGTGTGTG				8550
GCGACGACTT AGTCGTTATC TGIGAAAGTG CCGGGGTCCA GGAGGAOGCG				8600
GCGAGCTGTA GAGCTTCAAC CGAGGCTATG ACCAGGTACT CGCGCCCCCCC				8650
CGGGGACCCC CCACAACCAAG AATACGACTT GGAGCTTATA ACATCATGCT				8700
CCTCCAACGT GTCACTGCC CACGACGGGG CTGGAAAGAG GGCTCTACTAC				8750
CTTACCCCGTG ACCCTACAAAC CCGCTCGCG AGAGCGCGGT GGGAGACAGC				8800
AAGACACACT CCAGTCATT CCTGGCTAGG CAACATAATC ATGTTGGCC				8850
CCACACTGTG GCGGAGGATG ATACTGATGA CCACATTCTT TAGCGTCTTC				8900
ATAGCCAGGG ATCAGCTTGA ACAGGCTCTT AACTGTGAGA TCTAOGGAGC				8950
CTGCTACTCC ATAGAACCCAC TGGATCTACC TCCAATCATT CAAAGACTCC				9000
ATGGGCTCAAG CGCAATTTCAC CTCCACAGIT ACTCTCCAGG TGAAATCAAT				9050
AGGGTGGCCG CATGCGTCAG AAAACTGGG GTCCCCCGCT TCGGAGCTTG				9100
GAGACAOCCG GCGCGGAGCG TCGCGCTAG GCTTCTGTCC AGAGGGGCCA				9150
GGGCTGCGAT ATGIGGCAAG TACCTCTCA ACTGGGCGAT AAGAACAAAG				9200
CTCAAACCTCA CTCCAATAGC GCGCGCTGGC CGGCTGGACT TGTGGGGTIG				9250
GTTCACGGCT CGCTACAGCG CGGGAGACAT TTATCACAGC GTGCTCTCATG				9300
CGCGGCCCCG CTGGTCTGG TTTGCGCTAC TCTGCTCGC TGCAGGGGTA				9350
GGCATCTACC TCTCCCCAA CGATGAAAGG TTGGGGTAAA CACTCCGCC				9400
TCTTAAGCCA TTTCTGTTT TTTTTTTTTT TTTTTTTTTT TTTTTCTTTT				9450
TTTTTTCTT TCTTTCTT CTTCTTTTCC TTTCTTTTCA CCTCTTTAA				9500

FIG. 6E

H77C

10	20	30	40	50	
<u>1234567890</u>	<u>1234567890</u>	<u>1234567890</u>	<u>1234567890</u>	<u>1234567890</u>	
TCGTGGCTCC	ATCTTACGCC	TAGTCACGGC	TAGCTGIGAA	AGGTCCTGTGA	9550
GGCGCATGAC	TCCAGAGAGT	GCTGATACTG	GCCTCTCTGC	AGATCATGT	9599

H77C

10	20	30	40	50
1234567890	1234567890	1234567890	1234567890	1234567890
MSTINPKPQRK	TKRNINRRPQ	DVKFPGGGQI	VGGVYLLPRR	GPRLGVRATR
KTSERSQPRG	RRQPIPKARR	PEGRIWAQPG	YPWPLYGNEG	CGWAGWLLSP
RGSRPSWGPT	DPRRRSRNLG	KVIDTILTCGF	ADLMGYIPLV	GAPLOGAARA
LAHGVRVLED	GVNYATGNLP	GCSFSIFILLA	LLSCLTVPAS	AYQVRNSSL
YHVINDCPNS	SIVYEAADAI	LHTPGCVPCV	REGNASRCWV	AVITPIVATRD
GKLPTTQLRR	HIDLIVGSAT	LCSALYVGDL	CGSVFLMQL	FIFSPRRHWT
TQDNCSTYP	GHITGHRMAW	DMMNWSPTA	ALWVAQLLRI	PQAIMDMIAIG
AHWGVLAGIA	YFSMVGNWAK	VLMVLLLFAG	VDAEIHVTGG	NAGRITTAGLV
GLITPGAKQN	IQLININGSW	HINSTALNN	ESLNTGWL	LFYQHKFNSS
GCPERLASCR	RLTDFAQGNG	PISYANGSGL	DERPYCWHP	PRPOGIVPAK
SVCGPVYCFT	PSPWVVGTTD	RSGAPTYSWG	ANDIDVFVLN	NIRPPLGNWF
GCTWMNSTGF	TKVOGAPPCV	IGGVGNNTLL	CPIDCFRKHP	EATYSRCGSG
PWITPRCMVD	YPYRLWHYPC	TINYTIFKVR	MYVGGVEHRL	EAAONWIRGE
RCDLEDRDRS	EISPLLLSTT	QWQLPCSFT	TLPALSTGLI	HLHQNIVDVQ
YLYVGVGSSIA	SWAIKWEYVV	LLFLLLADAR	VCSCLWMMLL	ISQAEAALEN
LVLNAASLA	GHGGLVSFLV	FFCFAWYLKG	RWPGAVYAL	YGMWPLLLL
LALPORAYAL	DTEVAASCQGG	VVLVGLMALT	LSPYYKRYIS	WCMWLQYFL
TRVEAQLHW	VPPLNVRGGR	DAVILLMCVV	HPTLVFDITK	LLAIFGPLW
ILQASLLKVP	YFVRVQGLLR	ICALARKIAG	GHVQMAIIK	LGALTGTYVY
NHLTPLRWA	HNGLERLAVA	VEPVVFSRME	TKLITWGADT	AACGDIINGL
PVSARRGQEI	LLGPADGMVS	KGWRLLAPIT	AYAQQTTRGLL	GCITSLSLTGR
DKNQVEGEVQ	IVSTATQTFL	ATCINGVCWT	VYHGAGIRTI	ASPKGPVIQM
YTNDQDLVG	WPAFQGSRSL	TPCTCGSSDL	YLVTRHADVI	PVRRRGDSRG
SLLSPRPISY	LKGSSGGPLL	CPAGHAVGLF	RAAVCTRGA	KAVDFIPVEN
LGTIMRSPVF	TDNSSPPAVP	QSFQVAHLHA	PTGSGKSTKV	PAAYAAQGYK
VLVLNPSVAA	TLFGCAYMSK	AHGVDPNIRT	GVRTITIGSP	ITYSTYGF
ADGGCSGGAY	DIICDECHS	TDATSILGIG	TVLDQAETAG	ARLWVLATAT
PPGSVTVSHP	NIEEVALSTT	GEIPFYGKAI	PLEVIKGGRH	LIFCHSKKKC
DELAALKVAL	GINAVAYYRG	LDVSVIPTSG	DVVVSTDAL	MTGFTGDFDS
VIDCNCTVTQ	TVDFSLDPTF	TIEITTLQD	AVSRTQRRGR	TGRGKPGIYR
FVAPGERPSG	MFDSSVLCEC	YDAGCAWYEL	TPAETTVRLR	AYMNTPGLPV
CQDHLEFWEG	VFTGLTHIDA	HFLSQTKQSG	ENFPYLVAYQ	ATVCARAQAP
PPSWDQMWC	LIRLKPTLHG	PTPLLYRLGA	VQNEVILTHP	ITKYIMTCMS
ADLEWVTSIW	VLVGGVLAAL	AAACLSTGCV	VIVGRIVLSG	KPAIIPDREV
LYQEFDEMEE	CSQHLPYIEQ	GMMLAEQFKQ	KALGLIQTAS	RHAEVITPAV
QTNWQKLEVF	WAKHMWNFIS	GIQYLAGLST	LPGNPATIASL	MAFTAATISP
LTTGQTLFN	ILGGWVAAQL	AAPGAATAFV	GAGLAGAAIG	SVGLGKVLD
ILAGYAGVA	GALVAFKIMS	GEVPSTEDLV	NLLPATLSPG	ALVVGWCAA

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H77C

10	20	30	40	50
1234567890	1234567890	1234567890	1234567890	1234567890
ILRRHVGPGF GAVQAMNRLI AFASRGHNVS PIHYVPESDA AARVIAILSS				1950
LTVTQLLRLR HQWISSECIT PCSGWLRLDI WDWISEVLSDFKTIWLKAKLM				2000
PQLPGIPFVS CQGYRGWR GDGIMHTRCH CGAEITGHVK NGIMRIVGPR				2050
TCRNMSGTIF PINAYTTGPF TPLPAPNYKF ALWRVSAEY VEIRRVGDFH				2100
YVSGMTIDNL KCPOQIPSPE FFTELDGVRLLRFAPPCKPL LREEVSFRVG				2150
LHEYPVGSQF PCEPEPDVAV LTSMLTDPSH ITAEAAGRRL ARGSPPSMAS				2200
SSASQLSAPS LKATCTANHD SPDAAELIEAN LLWRQEMGGN ITRVESENKV				2250
VILDSEFDPLV AEEDEREVSV PAEILRKSR FARALPVWAR PDYNPPLVET				2300
WKKPDYEPPV VHGCPLPPPR SPPVPPPRKK RIVVLTESTIL STALAELATK				2350
SFGSSSTSGI TGDNITTSSE PAPSGCPPDS DVESYSSMPP LEGEPGDPDL				2400
SDGSWSIVSS GADTEDWCC SMSYSWTIGAL VTPCAAEEQK LPINALNSNL				2450
LRHHNLVYST TSRSAQQRQK KVTFDRLQVL DSHYQDVLKE VKAAASKVKA				2500
NLLSVEEACS LTPPHSAKSK FGYGAKDVRC HARKAVAHIN SWKDLLEDS				2550
VTPIDITIMA KNEVFCVQPE KGGRKPARLI VFPDLGVRVC EKMALYDWS				2600
KLPLAVMGSS YGFQYSPGQR VEFLVQAWKS KKTPMGFSYD TRCFDSLIVTE				2650
SDIRTEEAIIY QCCDLDPRQAR VAIKSLTERL YVGGPLINSR GENCGYRRCR				2700
ASGVLTTSOG NTILTGYIKAR AACRAAGLQD CIMLVOGDDL WVICESAGVQ				2750
EDAASLRAFT EAMTRYSAPP GDPPQPEYDL ELITSCSSNV SVAHDGAGKR				2800
VYLYLTRDPIT PLARAAWETA RHTPVNSWLGNIIIMFAPIIW ARMILMTHFF				2850
SVLIARDQLE QALNCETIYGA CYSIEPLDLP PIIQLRHGLS AFSLHSYSPG				2900
EINRVAACLR KLGVPPLRAW RHRARSVRAR LLSRGGRAAI CGKYLFWAV				2950
RTKLKLTPIA AAGRLLDLSGW FTAGYSGGDI YHSVSHARPR WFWFCLLLLIA				3000
AGVGIYLLPN R				3011

HC-J4

10	20	30	40	50
1234567890	1234567890	1234567890	1234567890	1234567890
GCCAGGGGGC	TGATGGGGGC	GACACTCAC	CATGAATCAC	TCCCCGTGGA
GGAACATACG	TCTTCAQGCA	GAAAGGGTCT	AGOCATGGCG	TTAGTATGAG
TGTGTTGGAG	CTTOCAGGAC	CCCCCTTCCC	GGGAGAGGCCA	TAGTGGTCTG
CGGAAACGGT	GAGTACACCG	GAATTGCCAG	GAOGACGGGG	TCCCTTCCTG
GATCAACCG	CTCAATGCC	GGAGATTGG	GGTGCCCCC	GGGAGACTGC
TAGCCGAGTA	GTTGTTGGTC	GGAAAGGCC	TTGTGGTACT	GGCTGATAGG
GTGCTTGCGA	GTGCCCCGGG	AGGTCTGTA	GAOGTGGCAC	CATGAGCACG
AATCCTAAAC	CTCAAAGAAA	AAACAAACGT	AAACACCAACC	GGGGCCCCACA
GGACGTCAAG	TTCCCCGGCG	GTGGTCAGAT	CGTGGTGGGA	GTTCACCTGT
TGCGCGCGAG	GGGGCCCCAGG	TTGGGTGTG	GGGGGACTAG	GAAGGCTTCC
GAGCGGTGCG	AACTCTGGG	AAGGGGACAA	OCTATCCAA	AGGCTCGCGG
AACCGAGGGC	AGGGCCCTGGG	CTCAGCCCCG	GTAACCTTGG	CCCCCTCTATG
GCAATGAGGG	CTTGGGTGCG	GCAGGATGSC	TCTGTCAAC	GGGGGGCTOC
CGGCGTAGIT	GGGGCCCCAC	GGACCCCCGG	CGTAGGTGCG	GTAACCTGGG
TAAGGTCACTC	GATAACCTTA	CATGGGGCTT	CGGGGATCTC	ATGGGGTACA
TTCCGCTCGT	GGGGGGGGGG	CTAGGGGGG	CTGCGAGGCG	CTTGGCACAC
GGTGTGGGG	TTCTGGAGGA	GGGGGTGAAAC	TATGCAACAG	GGAACTTGCC
CGGTTGCTCT	TTCTCTATCT	TCCTCTTGGC	TCTGCTGTCC	TGTTTGACCA
TOCCAGCTTC	CGCTTATGAA	GTGGGCAACG	TGTCCCCGAT	ATACCATGTC
ACGAACGACT	GCTCCAACTC	AAGCATTGIG	TATGAGGGCAG	CGGACGTGAT
CATGCATACT	CCCCGGTGCG	TGCCCCGTG	TCAGGAGGGT	AACAGCTCCC
GTGCTGGGT	AGCGCTCACT	CCCCGGCTCG	GGGGCAGGAA	TGCCAGGGTC
CCCACTACGA	CAATAQGAQG	CCACGTGCGAC	TTGCTCGTTG	GGACGGCTGC
TTTCTGCTCC	GCTATGTAQG	TGGGGGATCT	CTGCGGATCT	ATTTTCTCG
TCTCCGAGCT	GTTCACCTTC	TCGGCTCGCC	GGCATGAGAC	AGTGCAGGAC
TGCAACTGCT	CAATCTATCC	GGGCATGTA	TCAGGTCAAC	GCATGGCTTG
GGATATGATG	ATGAACCTGGT	CACCTACAAAC	AGGCGTAGTG	GTGTCGCACT
TGCTCCGGAT	CCCCAACGCT	GTGGTGGACA	TGGTGGGGGG	GGCCCACCTGG
GGAGGCTCTGG	GGGGGCTTGC	CTACTTATCC	ATGGTACGGA	ACTGGGCTAA
GGTTCGATT	GTGGGGCTAC	TCTTTCGCGG	CGTTGACGGG	GAGACCCACA
CGAOGGGGAG	GGTGGGGCGC	CACACCACT	CCCCGGTTCAC	GTCCCTTTTC
TCATCTGGGG	CGTCTCAGAA	AATCCAGCTT	GTGAATACCA	ACGGCAGCTG
GCACATCAAC	AGGACTGCCC	TAAATTGCAA	TGACTCCCTC	CAAACGGGT
TCTTTCGCGC	GCTGTTTAC	GCACACAAGT	TCAACTCGTC	GGGGTGGCCG
GAGGGCATGG	CCAGCTGCGG	CCCCATTGAC	GGGTTCGCCC	AGGGGTGGGG
CCCCATCACC	TATACTAACG	CTAACAGCTC	GGATCAGAGG	CCTTATTGCT
GGCATTACGC	GGCTCGACCG	TGTGGTGTG	TACCCCGCGTC	GCAGGTGTGT
GGTCCAGTGT	ATTGTTTAC	CCCCAGGCGT	GTGTTGGTGG	GGACCCACCGA

FIG. 7A

HC-J4

10	20	30	40	50
1234567890	1234567890	1234567890	1234567890	1234567890
TCGTTCCGGT	GTCCTTAOGT	ATAGCTGGGG	GGAGAATGAG	ACAGAOGTGA
TGCTCCTCAA	CAACACOGT	CGCGCACACAAG	GCAACTGGTT	CGGCTGTACA
TGGATGAATA	GTACTGGGTT	CACTAAGACG	TGCGGAGGTC	CGCGCTGTTAA
CATGGGGGGG	GTCGGTAACC	GCAACCTTGAT	CTGCCCCACG	GACTGCTTCC
GGAAGCACCC	CGAGGGCTACT	TACACAAAAT	GTGGCTGGGG	CGCGCTGGITG
ACACCTAGGT	GCCTAGTAGA	CTACCCATAC	AGGCTTTCGC	ACTAACCCCTG
CACTCTCAAT	TTTCCATCT	TTAAGGTTAG	GATGTATGTG	CGGGGGCGTGG
AGCACAGGCT	CAATGCGCA	TGCAATTGGA	CTCGAGGAGA	CGCGCTGTAAAC
TTGGAGGACA	GGGATAGGTC	AGAACTCAGC	CGCGCTGTGC	TGCTCTACAAC
AGAGTCCCAG	ATACTGCGCT	GTGCTTTCAC	CAACCTAACG	GCTTTATCCA
CTGGTTTGTAT	CCATCTOCAT	CAGAACATCG	TGGACGTGCA	ATACCTGTAC
GGTGTAGGGT	CAGCGTTGT	CTCGCTTGCA	ATCAAATGGG	AGTACATCCT
GTGCTTTTC	CTTCTCTGG	CAGACGCGCG	CGTGTGTGCC	TGCTTGTGGA
TGAATGCTGCT	GATAAGCCAG	GCTGAGGCCG	CGCTTAGAGAA	CTTGGTGGTC
CTCAATGCGG	CGTCCGTGCC	CGGAGCGCAT	GGTATTCTCT	CGCTTCTGT
GTTCCTCTGC	CGCGCTGGT	ACATTAAGGG	CAGGCTGGCT	CGTGGGGGGG
CGTATGCTTT	TTATGGGTA	TGGCGCTGC	TCCTGCTOCT	ACTGGCGTIA
CCACCAACGAG	CTTACCGCTT	GGACCGGGAG	ATGGCTGCAT	CGTGGGGGGG
TGCGGTTCTT	GTAGGCTCTG	TATTCTTGAC	CTTGTACCCA	TACTACAAAG
TGTTTCTCAC	TAGGCTCATA	TGGTGGTTAC	AATACTTTAT	CACCAGAGCC
GAGGGCGACA	TGCAAGTGTG	GGTCCCCCCC	CTCAACGTC	GGGGAGCCCG
CGATGCGCATC	ATCCTCTCA	CGTGTGGGT	TCATCCAGAG	TIAATTTTTG
ACATCACCAA	ACTCTGCTC	GOCATACTCG	GCCCCGCTCAT	CGTGTCTCCAG
GCTGGCATAA	CGAGAGTGCC	GTACTTGTG	CGCGCTCAAG	CGCTCATTCG
TCCATGCAATG	TTAGTGGGAA	AAGTGGCGG	GGGTCAATTAT	GFCCAAATGG
TCTTCATGAA	GCTGGGGCGG	CTGACAGGTA	CGTACGTTA	TAACCATCT
ACCCCACTGC	GGGACTGGGC	CCACCGGGGC	CTACGAGACC	TGCGGTGGC
GGTAGAGGCC	GTGCTCTCT	CGGCCATGGA	GACCAAGGTC	ATCAACTGGG
GAGCAGACAC	CGCTGGTGT	GGGGACATCA	TCTGGGTCT	ACCGCTCTCC
CCCCGAAGGG	GGAGGGAGAT	ATTTTGGGA	CGCGCTGATA	GTCTCGAAGG
GCAAGGGTGG	CGACTCCCTG	CGCCCATCAC	GGCGTACTCC	CAACAAACGC
GGGGCGTACT	TGGTTGCATC	ATCACTAGCC	TCACAGGCCG	GGACAAGAAC
CAGGTGGAAG	GGGAGGGTCA	AGTGGTTCT	ACCGCAACAC	AATCTTCT
GGCGACCTGC	ATCAACGGCG	TGTGCTGGAC	TCTCTCAT	CGCGCTGGCT
CGAAGACCT	AGCGGGTCCA	AAAGGTCCA	TCACCCAAAT	GTACACCAAT
GTAGACCTGG	ACCTCGTCGG	CTGGCAGGCG	CCCCCGGGGG	CGCGCTCCAT
GACACCATGC	AGCTGTGGCA	GCTOGGACCT	TTACTTGGTC	ACGAGACATG
CTGATGTCAT	TCCCGTGGC	CGCGGAGGCG	ACAGCAGGGG	AAGTCTACTC
				3800

FIG. 7B

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HC-J4

10	20	30	40	50
<u>1234567890</u>	<u>1234567890</u>	<u>1234567890</u>	<u>1234567890</u>	<u>1234567890</u>
TCGGGCGAGGC	CGGTCTCCGA	CCTGAAAGGC	TCTTGGGGTG	GTCATTGCT
TTGCCCTTTCG	GGGCAOGTGTG	TGGGCGCTTT	CGGCGCTGCT	GTGTGCGACCC
GGGGGGTGC	GAAGGGGGTG	GACITCATAC	CGGTGAGTC	TATGGAAACT
ACCATGCGGT	CTCGGGCTTT	CACAGACAAC	TCAAOCOCOC	CGGCTGTAC
GCAGACATTC	CAAGTGGCAC	ATCTGCAOGC	TCTTACTGGC	AGCGGCAAGA
GCACCAAAGT	CGCGCTGCG	TATGCAGOC	AAGGGTACAA	GGTGTGCGIC
CTGAACCGGT	CGGTGCGOOGC	CACCTTGGG	TTTGGGGGT	ATATGTCCAA
GGCACACGGT	ATCGACCGTA	ACATCAGAAC	TGGGTTAAGG	ACCATTACCA
CGGGCGGCCTC	CATTAOGTAC	TCCACCTATG	GCAAGTTCT	TGCGGAGGGT
GGCTGTTCTG	GGGGCGCTA	TGACATCATA	ATATGTGATG	AGTGCACACTC
AACTGACTCG	ACTAACATCT	TGGGCATOGG	CACAGTOCTG	GACCAAGGGG
AGACGGCTGG	AGCGGGGCTC	GTGCGCTCG	CCACCGCTAC	AOCTCOGGGA
TCGGTTACCG	TGCCACACCC	CAATATCGAG	GAAATAGGOC	TGTCACAAACAA
TGGAGAGATC	CCCTCTATG	GCAAAGOCAT	CCOCATTGAG	GOCATCAAGG
GGGGGAGGCA	TCTCAATTTC	TGOCATTCCA	AGAAGAAATG	TGACGAGCTC
GOOGCAAAGC	TGACAGGCGT	CGGACTGAAC	GCTGTAGGCAT	ATTACCGGGG
CCTTGTGTTG	TCCGTACATAC	CGCCTATOGG	AGACGTGCGT	GTGCGGGCAA
CAGACGCTCT	AATGACGGGT	TTCACCGGG	ATTTTGACTC	AGTGTATGAC
TGCAATACAT	GTGTCACCCA	GACAGTCGAC	TTCAGCTTGG	ATCCCCACCTT
CACCAATTGAG	ACGACGGAOG	TGCCCCAAGA	CGCGGTGCG	CGCTCGAAC
GGCGAGGTAG	AACTGGCAGG	GGTAGGAGTG	GCATCTACAG	GTITGTGACT
CCAGGAGAAC	GGCCCTCGGG	CATGTTGAT	TCTTCGGTCC	TGTGTGAGTG
CTATGACCGG	GGCTGTCCTT	GGTATGAGCT	CAOGCCCGCT	GAQACCTCGG
TTAGGTTCGG	GGCTTACCTA	AATACACCAG	GGTTCGGCGT	CTGCCAGGAC
CATCTGGAGT	TCTGGGAGAG	CGTCTTCACA	GGCCTCAACCC	ACATAGATCC
CCACTTCCIG	TOCCAGACTA	AACAGGGCAGG	AGACAACTTT	OCTTACCTTG
TGGCATATCA	AGCTACAGTG	TGCGGCCAGGG	CTCAAGCTCC	ACCTCCATOG
TGGGACCAAA	TGTGGAAGTG	TCTCATACGG	CTGAAACCTA	CACTGCAOGG
GCCAACACCC	CTGCTGTATA	GGCTAGGAGC	CGTCCAAAAT	GAGGTCACTCC
TCACACACCC	CATAACTAAA	TACATCATGG	CATGCATGTC	GGCTGACCTG
GAGGTGCTCA	CTAGCACCTG	GGTGTCTGGT	GGCGGAGTC	TTGCGCTTT
GGCGCGATAC	TGCCCTGAOGA	CAGGCAGTGT	GTCTTGTG	GGCAGGATCA
TCTTGTCCGG	GAAGCCAGCT	GTGCGTCCCG	ACAGGGAAAGT	CTCTCTACCA
GAGITCGATG	AGATGGAAGA	GTGTCCTCA	CAACTTCCTT	ACATCGAGCA
GGGAATCCAG	CTCGCCGAGC	AATTCAAGCA	AAAGGCCCTC	GGTTTGTGTC
AAACGGCACAC	CAAGCAAGCG	GAGGTGCTG	CTCCCGTGGT	GGAGTCACAG
TGGCGAGGCC	TTCAGACCTT	CTGGGCGAAG	CACATGTGGA	ATTTCATCAG
CGGAATACAG	TACCTAGCGAG	GCTTATOCAC	TCTGCCTGG	AAACCCCGCGA

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HC-J4

10	20	30	40	50
1234567890	1234567890	1234567890	1234567890	1234567890
TAGCATCATT	GATGGCATT	ACAGCTTCTA	TCACTAGGCCC	GCTCACCAAC
CAAAACACCC	TCCTGTTAA	CATCTTGGGG	GGATGGGTTGG	CTGCGCAACT
CGCTCTCCC	AGGGCTGGGT	CAGCTTGTG	GGGAGCCGGC	ATCGCGGGAG
CGGCTGTTGG	CAGCATAGGC	CTTGGGAAGG	TGCTTGTTGA	CATCTTGGCG
GGCTTAIGGGG	CAGGGGTAGC	CGGCGCACTC	GTGGCTTTA	AGGTCAITGAG
CGGCGAGGTG	CGCTCCACCG	AGGAOCTGGT	CAACTTACTC	OCTGOCATCC
TCTCTCTTGG	TGCGCTGGTC	GTGGGGGTGG	TGTGOGCAGC	AATACTGGGT
CGGCAOGTGG	GGGGGGGAGA	GGGGGCTGTG	CAGTGGATGA	AOCGGCTGAT
AGCGTTGGCT	TGAGGGGGTA	ACCAOGCTTC	CGCTTAOGCAC	TATGTGCGTG
AGAGCGAACG	TGCAGCAOGT	GTCACTCAGA	TCTCTCTCTAG	CGCTTAACCATC
ACTCAACTGC	TGAAGCGGCT	CCACCACTGG	ATTAATGAGG	ACTGCTCTAC
GCCATGCTCC	GGCTCGTGGC	TAAGGGATGT	TTGGGATTGG	ATATGCCACGG
TGTGACTGTA	CTTCAAGACC	TGGCTCCAGT	CGAAACTCT	GGCGCGGTTA
CGGGGAGTCC	CTTCTCTGTC	ATGCCAACCG	GGGTACAAGG	GAGTCCTGGCG
GGGGGACCGC	ATCATGCAAA	CCACCTGCC	ATGOGGGAGCA	CAGATCGCG
GACATGTCAA	AAACGGTTC	ATGAGGATCG	TAGGGCCTAG	AAACCTGCAGC
AACACGTGGC	ACGGAACGTT	CCCATCAAC	GCATACACCA	CGGGACCTTG
CACACCCCTCC	CCGGCGGCCA	ACTATTCCAG	GGCGCTATGG	CGGGTGGCTG
CTGAGGAGTA	CGTGGAGGTT	ACGGCGTGG	GGGATTTCGA	CTACGTGAGC
GGCATGACCA	CTGACAAOGT	AAAGTCCCCA	TGCCAGGTT	GGGGCCCCCGA
ATTCTTCACG	GAGGTGGATG	GAGTCGGGTT	GCACAGGTAC	GCTCGGGGT
GCAAACCTCT	TCTACGGGAG	GACGTCAOGT	TOCAGGTGG	GCTCAACCAA
TACCTGGTGT	GGTGGCAGCT	CCCATGCGAG	CCCGAACCGG	ACGTAACAGT
GCTTACTTCC	ATGCTCACCG	ATCCCTCCCA	CATTACAGCA	GAGACGGCTA
AGCGTAGGCT	GGCTAGAGGG	TCTCCCCCT	CTTAGCCAG	CTCATCAGCT
AGCCAGTTGT	CTGCGCCCTC	TTTGAAGGGG	ACATGCACTA	CCCAACCATGA
CTCCCCGGAC	CTGACCTCA	TCGAGGCGAA	CCTCTTGTGG	CGGCAGGAGA
TGGGGGGAAA	CATCACTCGC	GTGGAGTCAG	AGAATAAGGT	AGTAATTCTG
GACTCTTTCG	AACCGCTTCA	CGGGGAGGGG	GTGAGAGGG	AGATAATCCGT
CGGGGGCGGAG	ATCCCTGCGAA	AACTCAGGAA	GTCTCCCCCTCA	GGGTGCGCCA
TATGGGCACG	CCCGGACTAC	AACTCTCCAC	TGCTAGAGTC	CTGGAAAGGAC
CCGGACTACG	TCCCTCGGGT	GGTACAOGGA	TGCGCATTC	CACTAACCAA
GGCTCTCTCA	ATACCAOCTC	CAOGGAGAAA	GGGGAGGTT	GTCCTGACAG
AATCCAATGT	GTCTTCTGCC	TTGGGGGAGC	TOGOCACTAA	GAACCTTGGT
AGCTCCGGAT	CGTCGGCGGT	TGATAGCGGC	ACGGCGACCG	CCCTTCCCTGA
CCTGGCCCTCC	GAACGACGGTG	ACAAAGGATC	CGACGTTGAG	TGTTACTCCT
CCATGCCCGCC	CTTGAAGGGG	GAGGGGGGGG	ACCCCGATCT	CAGCGACGGG
TCTTGGTCTA	CGTGTAGTGA	GGAGGCTAGT	GAGGATGTGG	TCTGCTGCTC
				7600

FIG. 7D

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
AATGTCCTAT	AOGTGGACAG	GOGCCTGAT	CACGCCATGC	GCTGCGGAGG	7650
AAAGTAAGCT	GOCCATCAAC	COGTTGAGCA	ACTCTTTGCT	GGTCACCCAC	7700
AACATGGCT	ACGCCACAAC	ATCCCGCAGC	GCAAGCCIOC	GCCAGAAGAA	7750
GGTCACCTTT	GACAGATTGC	AAGTCTTGGA	TGATCATTAC	GGGGACGTAC	7800
TCAAGGAGAT	GAAGGGAAAG	GOGTCCACAG	TIAAGGCTAA	GCTTCTATCT	7850
ATAGAGGAGG	CCTGCAAGCT	GAOGCCCCCA	CATTGGCCA	AACTCAAATT	7900
TGGCTATCGG	GCAAAGGAOG	TOCGGAACCT	ATCCAGCAGG	GOOGTTAACCC	7950
ACATCCGCTC	CGTGTGGGAG	GACTTGCTGG	AAGACACTGA	AACACCAATT	8000
GACACCACCA	TCATGGCAAA	AAGTGGGTT	TTCCTGGCIOC	AACCAGAGAA	8050
GGGAGGGCGC	AAGCCAGCTC	GCCTTATCGT	ATTOCCAGAC	CTGGGAGTTTC	8100
GTGTATGCGA	GAAGATGGCC	CTTACGACG	TGGCTCCAC	OCTTCTCAG	8150
GCGGTGATGG	GCTOCTCATA	CGGATTCAA	TACTCCCCA	AGCAGGGGT	8200
CGAGTTCTG	GTGAATACT	GGAAATCAA	GAAATGCGCT	ATGGGCTTCT	8250
CATATGACAC	CGCTGTTTT	GACTCAAOGG	TCACTGAGAG	TGACATTGCT	8300
GTTGAGGAGT	CAATTIAOCA	ATGTTGTGAC	TTGGCCCCCG	AGGCCAGACAA	8350
GGCCATAAGG	TOGCTCACAG	AGCGGCTTTA	CATGGGGGT	CCCCCTGACTA	8400
ACTCAAAAGG	GCAGAACTGC	GGTTATGCC	GGTGGCGCGC	AAGTGGGTG	8450
CTGAOGACTA	GCTGCGTAA	TACOCTCACA	TGTTACTTGA	AGGCCACTGC	8500
AGCCTGTGCA	GCTGCAAAGC	TOCAGGACTG	CAOGATGCTC	GTGAACGGAG	8550
ACGACCTTGT	CGTTATCTGT	GAAGCCGGG	GAACCCAGGA	GGATGCGGGG	8600
GCGCTACGAG	OCTTCACGGA	GGCTATGACT	AGGTATTCCG	CCCCCCCCCGG	8650
GGATCCGCCC	CAACCAGAAAT	ACGACCTGGA	GCTGATAACA	TCATGTTCT	8700
CCAATGTGTC	AGTGGGGCAC	GATGCATCTG	GCAAAAGGGT	ATACIACCTC	8750
AACCGTGACC	CCACCAACCCC	CTTGCACGG	GCTGGTGGG	AGACAGCTAG	8800
ACACACTCCA	ATCAACTCTT	GGCTAGGCAA	TATCATCATG	TATGGCGCCA	8850
CCCTATGGGC	AAGGATGATT	CTGATGACTC	ACTTTTCTC	CATCTCTIA	8900
GCTCAAGAGC	AACTGAAAA	AGCCTGGAT	TGTCAGATCT	ACGGGGCTTG	8950
CTACTCCATT	GAGCCACTTG	ACCTAOCCTCA	GATCATTGAA	CGACTOCATG	9000
GTCTTAGGGC	ATTTACACTC	CACAGTTACT	CTCCAGGTGA	GATCAATAGG	9050
GTGGCTTCAT	GCCTCAGGAA	ACTTGGGGTA	CCACCCCTTG	GAACCTGGAG	9100
ACATCGGGCC	AGAAGTGTCC	GCGCTAAGCT	ACTGTCCCCAG	GGGGGGAGGG	9150
CGGCCACTTG	TGGCAGATAAC	CTCTTTAACT	GGGCAGTAAAG	GACCAAGCTT	9200
AAACTCACTC	CAATCCGGC	CGCGTCCCCAG	CTGGACTTGT	CTGGCTGGGT	9250
CGTGGCTGGT	TACAGGGGGG	GAGACATATA	TCACAGCTG	TCTCGTGGCC	9300
GACCCCGCTG	GTTCGGTGTG	TGCCTACTCC	TACTTTCTGT	AGGGGTAGGC	9350
ATTIACCTGC	TOCCCCAACCG	ATGAACGGGG	AGCTAAOCAC	TOCAGGCCCT	9400
AAGCCATTTC	CTGTTTTTTT	TTTTTTTTT	TTTTTTTTT	TCTTTTTTTT	9450
TTTCTTCTCT	TTCTTCTCTT	TTTCTCTT	TTTCTCCCTT	CTTAAATGGT	9500

FIG. 7E

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10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GGCTCCATCT	TAGCCCTAGT	CAOGGCTAGC	TGTGAAAGGT	CCGTGAGCG	9550
CATGACTGCA	GAGAGTGCTG	ATACTGGCT	CTCTGCCAGAT	CATGT	9595

10	20	30	40	50
<u>1234567890</u>	<u>1234567890</u>	<u>1234567890</u>	<u>1234567890</u>	<u>1234567890</u>
MSTINPKPQRK	TKRNINRRPQ	DVKFPGGGQI	VGGVYLLPRR	GPRLGVRAIR
KASERSQPRG	RRQPIPKARR	PEGRAWAQPG	YPWPLYGNNEG	LGNAGWLLSP
RGSRPSWGPT	DPRRRSRNLG	KVIDTILTOGF	ADLMGYIPLV	GAPLGGAAARA
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740	745	750
Glu Lys Leu Val Ile Leu His Ala Ala Ser Ala Ala Ser Cys Asn Gly		
755	760	765
Phe Leu Tyr Phe Val Ile Phe Phe Val Ala Ala Trp Tyr Ile Lys Gly		
770	775	780
Arg Val Val Pro Leu Ala Thr Tyr Ser Leu Thr Gly Leu Trp Ser Phe		
785	790	795
800		
Ser Leu Leu Leu Leu Ala Leu Pro Gln Gln Ala Tyr Ala Tyr Asp Ala		
805	810	815
Ser Val His Gly Gln Ile Gly Ala Ala Leu Leu Val Met Ile Thr Leu		
820	825	830
Phe Thr Leu Thr Pro Gly Tyr Lys Thr Leu Leu Ser Arg Phe Leu Trp		
835	840	845
Trp Leu Cys Tyr Leu Leu Thr Leu Gly Glu Ala Met Val Gln Glu Trp		
850	855	860
Ala Pro Pro Met Gln Val Arg Gly Gly Arg Asp Gly Ile Ile Trp Ala		
865	870	875
880		
Val Ala Ile Phe Tyr Pro Gly Val Val Phe Asp Ile Thr Lys Trp Leu		
885	890	895
Leu Ala Val Leu Gly Pro Ala Tyr Leu Leu Lys Gly Ala Leu Thr Arg		
900	905	910
Val Pro Tyr Phe Val Arg Ala His Ala Leu Leu Arg Met Cys Thr Met		
915	920	925
Ala Arg His Leu Ala Gly Gly Arg Tyr Val Gln Met Ala Leu Leu Ala		
930	935	940
Leu Gly Arg Trp Thr Gly Thr Tyr Ile Tyr Asp His Leu Thr Pro Met		

945	950	955	960
Ser Asp Trp Ala Ala Ser Gly Leu Arg Asp Leu Ala Val Ala Val Glu			
965	970	975	
Pro Ile Ile Phe Ser Pro Met Glu Lys Lys Val Ile Val Trp Gly Ala			
980	985	990	
Glu Thr Ala Ala Cys Gly Asp Ile Leu His Gly Leu Pro Val Ser Ala			
995	1000	1005	
Arg Leu Gly Arg Glu Val Leu Leu Gly Pro Ala Asp Gly Tyr Thr Ser			
1010	1015	1020	
Lys Gly Trp Ser Leu Leu Ala Pro Ile Thr Ala Tyr Ala Gln Gln Thr			
1025	1030	1035	1040
Arg Gly Leu Leu Gly Thr Ile Val Val Ser Met Thr Gly Arg Asp Lys			
1045	1050	1055	
Thr Glu Gln Ala Gly Glu Ile Gln Val Leu Ser Thr Val Thr Gln Ser			
1060	1065	1070	
Phe Leu Gly Thr Ser Ile Ser Gly Val Leu Trp Thr Val Tyr His Gly			
1075	1080	1085	
Ala Gly Asn Lys Thr Leu Ala Gly Ser Arg Gly Pro Val Thr Gln Met			
1090	1095	1100	
Tyr Ser Ser Ala Glu Gly Asp Leu Val Gly Trp Pro Ser Pro Pro Gly			
1105	1110	1115	1120
Thr Lys Ser Leu Glu Pro Cys Thr Cys Gly Ala Val Asp Leu Tyr Leu			
1125	1130	1135	
Val Thr Arg Asn Ala Asp Val Ile Pro Ala Arg Arg Gly Asp Lys			
1140	1145	1150	
Arg Gly Ala Leu Leu Ser Pro Arg Pro Leu Ser Thr Leu Lys Gly Ser			
1155	1160	1165	
Ser Gly Gly Pro Val Leu Cys Pro Arg Gly His Ala Val Gly Val Phe			
1170	1175	1180	
Arg Ala Ala Val Cys Ser Arg Gly Val Ala Lys Ser Ile Asp Phe Ile			
1185	1190	1195	1200
Pro Val Glu Thr Leu Asp Ile Val Thr Arg Ser Pro Thr Phe Ser Asp			

1205	1210	1215
Asn Ser Thr Pro Pro Ala Val Pro Gln Thr Tyr Gln Val Gly Tyr Leu		
1220	1225	1230
His Ala Pro Thr Gly Ser Gly Lys Ser Thr Lys Val Pro Val Ala Tyr		
1235	1240	1245
Ala Ala Gln Gly Tyr Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala		
1250	1255	1260
Thr Leu Gly Phe Gly Ala Tyr Leu Ser Lys Ala His Gly Ile Asn Pro		
1265	1270	1275
Asn Ile Arg Thr Gly Val Arg Thr Val Thr Thr Gly Ala Pro Ile Thr		
1285	1290	1295
Tyr Ser Thr Tyr Gly Lys Phe Leu Ala Asp Gly Gly Cys Ala Gly Gly		
1300	1305	1310
Ala Tyr Asp Ile Ile Cys Asp Glu Cys His Ala Val Asp Ser Thr		
1315	1320	1325
Thr Ile Leu Gly Ile Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly		
1330	1335	1340
Val Arg Leu Thr Val Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr		
1345	1350	1355
Thr Pro His Pro Asn Ile Glu Glu Val Ala Leu Gly Gln Glu Gly Glu		
1365	1370	1375
Ile Pro Phe Tyr Gly Arg Ala Ile Pro Leu Ser Tyr Ile Lys Gly Gly		
1380	1385	1390
Arg His Leu Ile Phe Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala		
1395	1400	1405
Ala Ala Leu Arg Gly Met Gly Leu Asn Ser Val Ala Tyr Tyr Arg Gly		
1410	1415	1420
Leu Asp Val Ser Val Ile Pro Thr Gln Gly Asp Val Val Val Val Ala		
1425	1430	1435
Thr Asp Ala Leu Met Thr Gly Tyr Thr Gly Asp Phe Asp Ser Val Ile		
1445	1450	1455
Asp Cys Asn Val Ala Val Thr Gln Val Val Asp Phe Ser Leu Asp Pro		

1460	1465	1470
Thr Phe Thr Ile Thr Thr Gln Ile Val Pro Gln Asp Ala Val Ser Arg		
1475	1480	1485
Ser Gln Arg Arg Gly Arg Thr Gly Arg Gly Arg Leu Gly Ile Tyr Arg		
1490	1495	1500
Tyr Val Ser Thr Gly Glu Arg Ala Ser Gly Met Phe Asp Ser Val Val		
1505	1510	1515
Leu Cys Glu Cys Tyr Asp Ala Gly Ala Ala Trp Tyr Glu Leu Thr Pro		
1525	1530	1535
Ser Glu Thr Thr Val Arg Leu Arg Ala Tyr Phe Asn Thr Pro Gly Leu		
1540	1545	1550
Pro Val Cys Gln Asp His Leu Glu Phe Trp Glu Ala Val Phe Thr Gly		
1555	1560	1565
Leu Thr His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly		
1570	1575	1580
Glu Asn Phe Ala Tyr Leu Thr Ala Tyr Gln Ala Thr Val Cys Ala Arg		
1585	1590	1595
Ala Lys Ala Pro Pro Pro Ser Trp Asp Val Met Trp Lys Cys Leu Thr		
1605	1610	1615
Arg Leu Lys Pro Thr Leu Val Gly Pro Thr Pro Leu Leu Tyr Arg Leu		
1620	1625	1630
Gly Ser Val Thr Asn Glu Val Thr Leu Thr His Pro Val Thr Lys Tyr		
1635	1640	1645
Ile Ala Thr Cys Met Gln Ala Asp Leu Glu Val Met Thr Ser Thr Trp		
1650	1655	1660
Val Leu Ala Gly Gly Val Leu Ala Ala Val Ala Ala Tyr Cys Leu Ala		
1665	1670	1675
Thr Gly Cys Val Cys Ile Ile Gly Arg Leu His Ile Asn Gln Arg Ala		
1685	1690	1695
Val Val Ala Pro Asp Lys Glu Val Leu Tyr Glu Ala Phe Asp Glu Met		
1700	1705	1710
Glu Glu Cys Ala Ser Arg Ala Ala Leu Ile Glu Glu Gly Gln Arg Ile		

1715	1720	1725
Ala Glu Met Leu Lys Ser Lys Ile Gln Gly Leu Leu Gln Gln Ala Ser		
1730	1735	1740
Lys Gln Ala Gln Asp Ile Gln Pro Thr Val Gln Ala Ser Trp Pro Lys		
1745	1750	1755
1760		
Val Glu Gln Phe Trp Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile		
1765	1770	1775
Gln Tyr Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Val Ala		
1780	1785	1790
Ser Met Met Ala Phe Ser Ala Ala Leu Thr Ser Pro Leu Ser Thr Ser		
1795	1800	1805
Thr Thr Ile Leu Leu Asn Ile Leu Gly Gly Trp Leu Ala Ser Gln Ile		
1810	1815	1820
Ala Pro Pro Ala Gly Ala Thr Gly Phe Val Val Ser Gly Leu Val Gly		
1825	1830	1835
1840		
Ala Ala Val Gly Ser Ile Gly Leu Gly Lys Val Leu Val Asp Ile Leu		
1845	1850	1855
Ala Gly Tyr Gly Ala Gly Ile Ser Gly Ala Leu Val Ala Phe Lys Ile		
1860	1865	1870
Met Ser Gly Glu Lys Pro Ser Met Glu Asp Val Val Asn Leu Leu Pro		
1875	1880	1885
Gly Ile Leu Ser Pro Gly Ala Leu Val Val Gly Val Ile Cys Ala Ala		
1890	1895	1900
Ile Leu Arg Arg His Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met		
1905	1910	1915
1920		
Asn Arg Leu Ile Ala Phe Ala Ser Arg Gly Asn His Val Ala Pro Thr		
1925	1930	1935
His Tyr Val Thr Glu Ser Asp Ala Ser Gln Arg Val Thr Gln Leu Leu		
1940	1945	1950
Gly Ser Leu Thr Ile Thr Ser Leu Leu Arg Arg Leu His Asn Trp Ile		
1955	1960	1965
Thr Glu Asp Cys Pro Ile Pro Cys Gly Ser Trp Leu Arg Asp Val		

1970	1975	1980
Trp Asp Trp Val Cys Thr Ile Leu Thr Asp Phe Lys Asn Trp Leu Thr		
1985	1990	1995
Ser Lys Leu Phe Pro Lys Met Pro Gly Leu Pro Phe Val Ser Cys Gln		
2005	2010	2015
Lys Gly Tyr Lys Gly Val Trp Ala Gly Thr Gly Ile Met Thr Thr Arg		
2020	2025	2030
Cys Pro Cys Gly Ala Asn Ile Ser Gly Asn Val Arg Leu Gly Ser Met		
2035	2040	2045
Arg Ile Thr Gly Pro Lys Thr Cys Met Asn Ile Trp Gln Gly Thr Phe		
2050	2055	2060
Pro Ile Asn Cys Tyr Thr Glu Gly Gln Cys Val Pro Lys Pro Ala Pro		
2065	2070	2075
Asn Phe Lys Val Ala Ile Trp Arg Val Ala Ala Ser Glu Tyr Ala Glu		
2085	2090	2095
Val Thr Gln His Gly Ser Tyr His Tyr Ile Thr Gly Leu Thr Thr Asp		
2100	2105	2110
Asn Leu Lys Val Pro Cys Gln Leu Pro Ser Pro Glu Phe Phe Ser Trp		
2115	2120	2125
Val Asp Gly Val Gln Ile His Arg Phe Ala Pro Thr Pro Lys Pro Phe		
2130	2135	2140
Phe Arg Asp Glu Val Ser Phe Cys Val Gly Leu Asn Ser Phe Val Val		
2145	2150	2155
Gly Ser Gln Leu Pro Cys Asp Pro Glu Pro Asp Thr Asp Val Leu Met		
2165	2170	2175
Ser Met Leu Thr Asp Pro Ser His Ile Thr Ala Glu Thr Ala Ala Arg		
2180	2185	2190
Arg Leu Ala Arg Gly Ser Pro Pro Ser Glu Ala Ser Ser Ser Ala Ser		
2195	2200	2205
Gln Leu Ser Ala Pro Ser Leu Arg Ala Thr Cys Thr Thr His Gly Lys		
2210	2215	2220
Ala Tyr Asp Val Asp Met Val Asp Ala Asn Leu Phe Met Gly Gly Asp		

2225	2230	2235	2240
Val Thr Arg Ile Glu Ser Gly Ser Lys Val Val Val Leu Asp Ser Leu			
2245	2250	2255	
Asp Pro Met Val Glu Glu Arg Ser Asp Leu Glu Pro Ser Ile Pro Ser			
2260	2265	2270	
Glu Tyr Met Leu Pro Lys Lys Arg Phe Pro Pro Ala Leu Pro Ala Trp			
2275	2280	2285	
Ala Arg Pro Asp Tyr Asn Pro Pro Leu Val Glu Ser Trp Lys Arg Pro			
2290	2295	2300	
Asp Tyr Gln Pro Ala Thr Val Ala Gly Cys Ala Leu Pro Pro Pro Arg			
2305	2310	2315	2320
Lys Thr Pro Thr Pro Pro Arg Arg Arg Arg Thr Val Gly Leu Ser			
2325	2330	2335	
Glu Asp Ser Ile Gly Asp Ala Leu Gln Gln Leu Ala Ile Lys Ser Phe			
2340	2345	2350	
Gly Gln Pro Pro Pro Ser Gly Asp Ser Gly Leu Ser Thr Gly Ala Gly			
2355	2360	2365	
Ala Ala Asp Ser Gly Ser Gln Thr Pro Pro Asp Glu Leu Ala Leu Ser			
2370	2375	2380	
Glu Thr Gly Ser Ile Ser Ser Met Pro Pro Leu Glu Gly Glu Leu Gly			
2385	2390	2395	2400
Asp Pro Asp Leu Glu Pro Glu Gln Val Glu Pro Gln Pro Pro Pro Gln			
2405	2410	2415	
Gly Gly Val Ala Ala Pro Gly Ser Asp Ser Gly Ser Trp Ser Thr Cys			
2420	2425	2430	
Ser Glu Glu Asp Asp Ser Val Val Cys Cys Ser Met Ser Tyr Ser Trp			
2435	2440	2445	
Thr Gly Ala Leu Ile Thr Pro Cys Ser Pro Glu Glu Lys Leu Pro			
2450	2455	2460	
Ile Asn Pro Leu Ser Asn Ser Leu Leu Arg Tyr His Asn Lys Val Tyr			
2465	2470	2475	2480
Cys Thr Thr Thr Lys Ser Ala Ser Leu Arg Ala Lys Lys Val Thr Phe			

2485	2490	2495
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Asp Arg Met Gln Val Leu Asp Ser Tyr Tyr Asp Ser Val Leu Lys Asp	2500	2505
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Ile Lys Leu Ala Ala Ser Lys Val Thr Ala Arg Leu Leu Thr Met Glu	2515	2520
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Glu Ala Cys Gln Leu Thr Pro Pro His Ser Ala Arg Ser Lys Tyr Gly	2530	2535
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Phe Gly Ala Lys Glu Val Arg Ser Leu Ser Gly Arg Ala Val Asn His	2545	2550
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Ile Lys Ser Val Trp Lys Asp Leu Leu Glu Asp Ser Glu Thr Pro Ile	2565	2570
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Pro Thr Thr Ile Met Ala Lys Asn Glu Val Phe Cys Val Asp Pro Thr	2580	2585
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Lys Gly Gly Lys Lys Ala Ala Arg Leu Ile Val Tyr Pro Asp Leu Gly	2595	2600
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Val Arg Val Cys Glu Lys Met Ala Leu Tyr Asp Ile Thr Gln Lys Leu	2610	2615
---	------	------

Pro Gln Ala Val Met Gly Ala Ser Tyr Gly Phe Gln Tyr Ser Pro Ala	2625	2630
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Gln Arg Val Glu Phe Leu Leu Lys Ala Trp Ala Glu Lys Lys Asp Pro	2645	2650
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Met Gly Phe Ser Tyr Asp Thr Arg Cys Phe Asp Ser Thr Val Thr Glu	2660	2665
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Arg Asp Ile Arg Thr Glu Glu Ser Ile Tyr Arg Ala Cys Ser Leu Pro	2675	2680
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Glu Glu Ala His Thr Ala Ile His Ser Leu Thr Glu Arg Leu Tyr Val	2690	2695
---	------	------

Gly Gly Pro Met Phe Asn Ser Lys Gly Gln Thr Cys Gly Tyr Arg Arg	2705	2710
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Cys Arg Ala Ser Gly Val Leu Thr Thr Ser Met Gly Asn Thr Ile Thr	2725	2730
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Cys Tyr Val Lys Ala Leu Ala Ala Cys Lys Ala Ala Gly Ile Ile Ala

2740

2745

2750

Pro Thr Met Leu Val Cys Gly Asp Asp Leu Val Val Ile Ser Glu Ser
2755 2760 2765

Gln Gly Thr Glu Glu Asp Glu Arg Asn Leu Arg Ala Phe Thr Glu Ala
2770 2775 2780

Met Thr Arg Tyr Ser Ala Pro Pro Gly Asp Pro Pro Arg Pro Glu Tyr
2785 2790 2795 2800

Asp Leu Glu Leu Ile Thr Ser Cys Ser Ser Asn Val Ser Val Ala Leu
2805 2810 2815

Gly Pro Gln Gly Arg Arg Arg Tyr Tyr Leu Thr Arg Asp Pro Thr Thr
2820 2825 2830

Pro Ile Ala Arg Ala Ala Trp Glu Thr Val Arg His Ser Pro Val Asn
2835 2840 2845

Ser Trp Leu Gly Asn Ile Ile Gln Tyr Ala Pro Thr Ile Trp Ala Arg
2850 2855 2860

Met Val Leu Met Thr His Phe Phe Ser Ile Leu Met Ala Gln Asp Thr
2865 2870 2875 2880

Leu Asp Gln Asn Leu Asn Phe Glu Met Tyr Gly Ala Val Tyr Ser Val
2885 2890 2895

Ser Pro Leu Asp Leu Pro Ala Ile Ile Glu Arg Leu His Gly Leu Asp
2900 2905 2910

Ala Phe Ser Leu His Thr Tyr Thr Pro His Glu Leu Thr Arg Val Ala
2915 2920 2925

Ser Ala Leu Arg Lys Leu Gly Ala Pro Pro Leu Arg Ala Trp Lys Ser
2930 2935 2940

Arg Ala Arg Ala Val Arg Ala Ser Leu Ile Ser Arg Gly Gly Arg Ala
2945 2950 2955 2960

Ala Val Cys Gly Arg Tyr Leu Phe Asn Trp Ala Val Lys Thr Lys Leu
2965 2970 2975

Lys Leu Thr Pro Leu Pro Glu Ala Arg Leu Leu Asp Leu Ser Ser Trp
2980 2985 2990

Phe Thr Val Gly Ala Gly Gly Asp Ile Tyr His Ser Val Ser Arg

2995

3000

3005

Ala Arg Pro Arg Leu Leu Leu Phe Gly Leu Leu Leu Leu Phe Val Gly
3010 3015 3020

Val Gly Leu Phe Leu Leu Pro Ala Arg
3025 3030

INTERNATIONAL SEARCH REPORT

Internal Application No

PCT/US 00/15293

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/51 C07K14/18 C12Q1/68 C12N7/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, BIOSIS, MEDLINE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>WO 95 21922 A (PILOT MATIAS TAMI J ;BUIJK SHERI L (US); SIMONS JOHN N (US); ABBOT) 17 August 1995 (1995-08-17) page 4, line 18 -page 6, line 17 page 55, line 24 -page 56, line 19 page 76; example 5 page 89, line 18 -page 96 page 109; example 15 page 148; example 21 page 427, line 17 -page 432 claims ---- -/-</p>	1,2,4-18



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

° Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

Date of mailing of the international search report

17 October 2000

31/10/2000

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Authorized officer

Andres, S

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/15293

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	SCARSELLI ELISA ET AL: "GB virus B and hepatitis C virus NS3 serine proteases share substrate specificity." JOURNAL OF VIROLOGY, vol. 71, no. 7, July 1997 (1997-07), pages 4985-4989, XP002150190 ISSN: 0022-538X cited in the application the whole document ---	19,24-26
A	HONDA MASA0 ET AL: "A phylogenetically conserved stem-loop structure at the 5' border of the internal ribosome entry site of hepatitis C virus is required for cap-independent viral translation." JOURNAL OF VIROLOGY, vol. 73, no. 2, February 1999 (1999-02), pages 1165-1174, XP002150191 ISSN: 0022-538X cited in the application the whole document ---	19,22,23
A	YANAGI MASAYUKI ET AL: "In vivo analysis of the 3' untranslated region of the hepatitis C virus after in vitro mutagenesis of an infectious cDNA clone." PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES, vol. 96, no. 5, 2 March 1999 (1999-03-02), pages 2291-2295, XP002150192 ISSN: 0027-8424 cited in the application ---	
A	YANAGI M ET AL: "Transcripts of a chimeric cDNA clone of hepatitis C virus genotype 1b are infectious in vivo" VIROLOGY, vol. 244, no. 1, 1998, pages 161-172, XP002089701 ISSN: 0042-6822 cited in the application ---	
P,X	BUKH JENS ET AL: "Toward a surrogate model for hepatitis C virus: An infectious molecular clone of the GB virus-B hepatitis agent." VIROLOGY, vol. 262, no. 2, 30 September 1999 (1999-09-30), pages 470-478, XP002150193 ISSN: 0042-6822 the whole document --- -/-	1-16,19

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/15293

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	SBARDELLATI ANDREA ET AL: "Identification of a novel sequence at the 3' end of the GB virus B genome." JOURNAL OF VIROLOGY, vol. 73, no. 12, December 1999 (1999-12), pages 10546-10550, XP002150194 ISSN: 0022-538X the whole document -----	1-16, 19
P,X	BUTKIEWICZ N. ET AL.: "Virus-specific cofactor requirement and chimeric hepatitis C virus/GB virus B nonstructural protein 3." J VIROL 2000 MAY;74(9):4291-301, XP002150195 the whole document -----	19, 24-26, 33-35, 37, 39

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 00/15293

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
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